

AL-TAHADI UNIVERSITY FACULTY OF SCIENCE DEPARTMENT OF ZOOLOGY

PREVALENCE AND MORPHOLOGICAL STUDIES ON THE LARVAL CYSTS OF ECHINOCOCCUS GRANULOSUS (BATSCH, 1786) IN HERBIVORUS ANIMALS IN SIRT-LIBYA

A thesis submitted in partial fulfillment of the requirements for the degree of Master of Science.

By
Abdelkader Mohammed Abdelkader Muftah

Supervisor Dr. Hamed H. Kassem

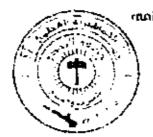
Associate Professor of Parasitology Faculty of Science, Garyounis University Benghazi, Libya

December - 2006

الرفقة بالأقباب فيادان بخرياتها G. S. P. L. A. J.

AL TAHDI UNIVERSITY

الرقع الإشابي، لي كل 26/ 1/ 2007 في



الصماهرية المرية الليارة الشميية الإشاراكية نامجاميا جاممة النجدي

كبلية العلوم

المالة 14 / 2007 ف

Faculty of Science Department of Biology Title of Thesis

((Prevalence, Morphological Studies on the Larval Cysts of Echinococcus Granulosue (Batsch, 1786) in Herbivorus Animals in Sirt – Libya))

By

Abdelkader Mohammed Abdelkader Muftah

Approved by:

Dr. Hamed H. Kassem (Supervisor)

Dr. Abdunnaser Ali El-Buni (External examiner)

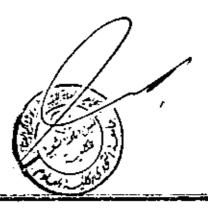
Dr. Fahima Hamed Hamam (Internal examiner)

Countersigned by:

Dr. Mohamed Ali Salem

(Dean of Faculty of Science)

4/12/06 - 10-





شَرِّنَ ﴿ لَانْ الْمُؤْكِدُ الْمُؤْكِدُ } النَّرِينَ * فَلَهُمْ عَالَحُهُمْ } سورة الإسراء – الأية 85

CONTENTS

Page
Contents
Acknowledgement III
List of Tables IV
List of FiguresVII
List of Plates IX
1. Introduction
2. Literature Review 3
2.1 Morphological characters of E. granulosus
2.2 Life cycle patterns of E. granulosus in Libya 4
2.3 Strains of E. granulosus
2.4 Biochemical profile of hydatid cysts 8
2.5 Epidemiology of hydatidosis 8
2.6 Hydatidosis in Libya
2.6.1 Human hydatidosis
2.6.2 Livestock hydatidosis
2.6.3 Definitive host Hydatidosis
3. Materials and Methods
3.1. study area
3.2. Animals
3.2.1. Studied animals
3.3. The abattoirs 24
3.4 Detection of parasite 24
3.5 Examination of hydatid cysts
3.6 Determination of hydatid cyst characteristics
3.6.1 Cyst size

ACKNOWLEDGEMENTS

Thanks to Allah for giving me the efforts to complete this work. My sincere gratitude and appreciation to my supervisor Dr. Hamed H. Kussem. Associate Professor of Parasitology, Zoology Department, Faculty of Science. Garyounis University, for suggesting, planning the point of research, supervising the work, reading the manuscript, his kind encouragement and concern for my welfare.

Sincere appreciation and best regards should be expressed to the University of Al-Tahadi, Faculty of Science and Zoology Department in providing all the required facilities for graduate studies. A specific acknowledgement to Dr. Mohamed Al-Fergani, Dean of the Faculty of Science, Al-Tahadi University and Dr. Ahmed Al-Mahgoub, Head of Zoology Department, Al-Tahadi University for their help and applying the necessary facilities required for this work.

I would like to extend my thanks to the veterinarians and inspectors at Sirt abattoir for their timely help regarded to collection samples for this study.

I am most grateful to the Department of Zoology, Faculty of Science, Garyounis University in which part of this work had been done.

Lastly a word of appreciation and sincere thanks to my brother Abdalsalam, for his assistance and moral support and to may parents, wife for their help and encouragement throughout the period of my study.

LIST OF TABLES

Page
Table 1 : Number of rostellum hooks from liver and lungs
hydatid cysts of Echinococcus granulosus examined
per species
Table 2: Overall prevalence of hydatid cysts in slaughtered
animals hosts from Sirt
Table 3: Prevalence of hydatid cysts in slaughtered
animals according to their sex
Table 4: Prevalence of hydatid cysts in slaughtered animals
according to their age41
Table 5: Prevalence of hydatid cysts in slaughtered animals
according to their seasons
Table 6: Location of hydatid cysts in liver and lungs of
sheep, goats, cattle and camels
Table 7: Location of hydatid cysts in liver and lungs of sheep,
goats, cattle and camels according to their age
Table 8: Location of hydatid cysts in liver and lungs of sheep,
goats, cattle and camels according to their sex
Table 9: Intensity of infection in liver and lungs of sheep,
goats, cattle and camels

Table 10: Intensity of infection in sheep, goats, cattle and camels according to their age
camets according to their ago
Table 11: Intensity of infection in sheep, goats, cattle and
camels according to their sex
Table 12: The size of hydatid cysts in liver and lungs of
sheep, goats, cattle and camels
Table 13: The size of hydatid cysts of sheep, goats, cattle and
camels according to their sex
Table 14: The size of hydatid cysts of sheep, goats, cattle and
camels according to their age
Table 15: Classification of hydatid cysts of examined sheep,
goats, cattle and camels according to their fertility
and physical contents
Table 16: The fertility of hydatid cyst of sheep, goats, cattle and
camels according to their age 63
Table 17: The fertility of hydatid cysts of sheep, goats and
camels according to their sex
Table 18: Comparsion between the fertility of hydatid cysts in liver
and lungs of sheep, goats and camels

Table 19: Comparsion between the viability of hydatid cyst protoscolices from liver, lungs of sheep, goats and camels 64
protosconces from tiver, langs of sheep, godio and the
Table 20: The viability of cyst protoscoleces of examined sheep, goats and camels according to their age
Table 21: The viability of cyst protoscoleces of sheep, goats and camels according to their sex
Table 22: The viability of hydatid cyst protoscoleces in liver and lungs of sheep, goats and camels
Table 23: The viability of cyst protoscoleces of examined sheep, goats and camels according to the cyst size
Table 24: Measurement of protoscoleces and total number of their hooks from liver and lungs of examined sheep, goats and camels
Table 25: Measurements of rostellum hooks from hydatid cysts of sheep, goats and camels

LIST OF FIGURES

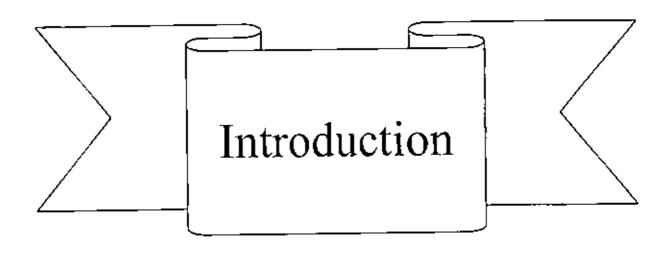
Page
Figure 1: Diagrammatic representation of the life cycle of
E. granulosus indicate sheep-dog life cycle
Figure 2: Map of Libya-showing place of the study (Sirt city) 20
Figure 3: Overall prevalence of hydatid cysts in slaughtered sheep,
goats, cattle and camels in Sirt
Figure 4: Prevalence of hydatid cysts in slaughtered according to their sex
Figure 5: Prevalence of hydatidosis in sheep at different age
groups 43
Figure 6: Prevalence of hydatidosis in goats at different age groups
Figure 7: Prevalence of hydatidosis in cattle at different age groups
Figure 8: Prevalence of hydatidosis in camels at different age groups
Figure 9: Prevalence of hydatidosis in sheep at different seasons 44
Figure 10: Prevalence of hydatidosis in goats at different seasons 44

Figure 11: Prevalence of hydatidosis in cattle at different seasons	. 44
Figure 12: Prevalence of hydatidosis in camels at different seasons	. 44
Figure 13: Location of hydatid cysts in liver and lungs of sheep, goats, cattle and camels	. 47
Figure 14: Intensity of infection sheep, goats, cattle and camels	50
Figure 15: Fertility of hydatid cysts from examined sheep, goats, cattle and camels	62

LIST OF PLATES

Page
Plate 1a, b: Libyan Bar bary sheep (Ovis aries)
Plate 2: Goats (Capra hircus)
Plate 3: Libyan camels (Camelus dromedaries)
Plate 4: Cattle (Bos taurus) breads (Friesian) (a), local (b) 23
Plate 5: Sheep hall of Sirt abattoir showing manual processing of carcasses
Plate 6: Slaughtered sheep investigated by abattoir veterinarian to find out any hydatid cysts
Plate 7: Inspection of sheep liver for hydatid cysts
Plate 8: Examination of livers and collection of hydatid cysts
Plate 9: Microscopic examination of hydatid cysts and morphology of protoscoleces and rostellum in fertile cysts
Plate 10: Hydatid cysts showing viable (v) and non-viable (nv) protoscoleces following treatment with 0.1 % eosin to test viability (X = 10)
Plate 11: Hydatid cyst from sheep liver, broad capsule showing viable and non-viable protoscoleces (X = 40)

Plate 12: Protoscolece from sheep liver hydatid cysts
showing the arrangement of rostellum hooks
(X = 100) 31
Plate 13: The method by which the proper disposal
of condemned organs
Plate 14: Dogs (The main definitive host) near the place
where the disposal of condemned organs
Plate 15: Showing the dog-definitive host feeding on
dead camel
Plate 16: Sirt dump-Note the dogs feeding on disposal
of condemned organs
Plate 17a , b : Hydatid cysts of Echinococcus granulosus
in camel liver 54
Plate 18 a , b : Hydatid cysts of Echinococcus granulosus
in cattle liver 55
Plate 19: Sheep liver infected with hydatid cysts
Plate 20: Goat lungs infected with hydatid cysts
Plate 21: Rostellum hooks from cyst showing measurement
recorded X = 100 (a) Total length, (b) Blade length,
(c) Handle length, (d) Hook width 70



1. INTRODUCTION

Hydatidosis is a widespread zoonotic disease infecting large number of animals and humans (Bource, 2001). It is caused by the larval stage for dogs and by eggs for humans and animals of any one of the four species of genus *Echinococcus* Rudolphi, 1801. Those four species are *Echinococcus granulosus* (Batsch, 1786), *Echinococcus multilocularis* (Leuckart, 1863), *Echinococcus vogeli* (Rausch and Bernstein, 1972) and *Echinococcus oligarthrus* (Diesing, 1963) (Thompson and Lymbery, 1988). The latter two species are rare infections are confined to Central and South America. The taxonomy of four species of genus *Echinococcus* was based on (1) morphological variations alone and (2) mitochondrial DNA sequencing showed these species to be genetically distinct (Bowles *et al.*, 1992). The life cycle patterns and distribution of these four species was reviewed by Rausch (1995).

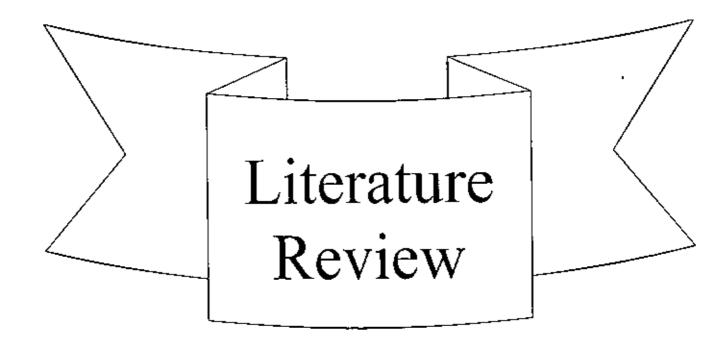
Adult Echinococcus spp. are small, true tapeworms belonging to the class Cestoda within the phylum platyhelminths. Subfamily Echinococcinae (Thompson, 1995; Rausch, 1997). They have a characteristic features of order cyclophyllidea. Echinococcus spp. are obligatory parasites of mammals requiring two hosts to complete their life cycle. Both adult and larval stages of these species are morphologically and biologically dissimilar.

Hydatidosis has been reported in both urban and rural communities where dog, the main definitive host, have been access to raw offal through home-slaughter from poorly regulated abattoirs or from scavenging carcasses or discarded offal (Watson-Jones *et al.*, 1997). Man can be infected by ingesting eggs from canine faeces on vegetables

or fruits or from handling dogs (Onah et al., 1989). In addition to economic importance, hydatidosis is a great threat to public health and many human cases require surgical interference (Haridy et al., 2000).

The prevalence rates of cystic hydatidosis in livestock are indicators of environmental transmission and potential risk for human (Ibrahem and Craig, 1998). Hydatid disease is endemic in many animals raising countries particularly in Middle East, Mediterranean (FAO, 1993; Clavel et al., 1999 and Erman et al., 2001), North Africa (Matossian et al., 1977 and Gebreel et al., 1983). Hydatid disease appears to be endemic in Libya (Gebreel et al., 1983). However little work has been published in Libya (Dar and Taguri, 1978; Gebreel et al., 1983; Kalani et al., 1984; Gusbi, 1987; Gusbi et al., 1987; Awan et al., 1990; Shambesh et al., 1992, 1999; Khan and Kidwai, 1996; Khan and El-Buni, 1999; Tashani et al., 2002 and Mohamed et al., 2004).

The aim of this study are (1) Determining the current prevalence of cystic echinococcosis in various domestic herbivorous animals slaughtered for human consumption in Sirt. (2) Study the morphological characters of larval stage of *Echinococcus granulosus* in these animals. (3) Analyse the different factors which may responsible for transmission and spread of the disease in Sirt. and (4) Analysis of obtained results using different statistical methods.



2. LITERATURE REVIEW

2.1 Morphological characters of \boldsymbol{E} , $\boldsymbol{granulosus}$:

Adult worm: The adult parasites in the intestine of dogs represent one of the smallest tapeworms, with varying length between 3 and 9 mm and usually of only 3-5 proglottids, the scolex is globular in shape and has a prominent rostellum, armed with a double row of between 30 and 36 hooks (Saidi, 1976; Gutierrez, 1990 and Thompson and Mc Manus, 2002). The eggs are very similar to those of the genus *Taenia*, and measure between 30 and 40 µm in diameter. The eggs are resistant to temperature ranging from 30 to 38 °C (Gemmel and Lawson, 1986; Thompson *et al.*, 1995). An infected dog is known to be capable of sheeding about 120 million eggs per week (Morris and Richards, 1992).

The metacestode (hydatids): The term metacestode and hydatid cysts are used to describe the larval stage of all species of *Echinococcus*. Clinical and economical aspect of infection due to this stage (Thompson, 1995). The hydatid cysts are large, roughly spherical, fluid filled hollow bladders, containing numerous protoscoleces (forming the so-called hydatid sand), broad capsules and daughter cysts which are identical in form to their parent cyst. The cyst wall itself consists of an outer laminated hyaline wall, supporting the whole cyst. Beneath this there is a nucleated germinal layer, sudded with developing broad capsules, which may eventually break of to float freely in the fluid filled cyst. The protoscoleces are formed within the broad capsules, which may rupture to give the free protoscoleces in the hydatid fluid (Morris and Richards, 1992 and Muller, 2002).

2.2 Life cycle patterns of E. granulosus in Libya:

The life cycle of *E. granulosus* a domestic cycle (Fig. 1), involving domestic dog as the definitive host for the adult worms and various species of livestock including sheep, goats, cattle, camels, buffaloes, pigs, horses and donkeys as intermediate hosts for the larval stage (El-Kordy, 1946; Dar and Taguri, 1978, 1979; Larbaoui *et al.*, 1980; Gusbi *et al.*, 1987; Pandey *et al.*, 1988; Ahmed, 1991).

Dogs are responsible for the contamination of the pasture with the infective eggs (Gusbi, 1987). Various domesticated animals harbors the larval hydatid cyst stage (Gusbi et al., 1990). Human infection may acquired through the ingestion of eggs with vegetable or drinking water or through direct contact with dogs as in case of children (Gebreel et al., 1983 and Shambesh et al. 1992) or through tending the herds of goats and sheep (Kalani et al., 1984). Fossati (1970) reported that human infection may acquired through inhalation of dust containing eggs, this may occurs because Libyan climate characteristic with "gibli" storm can assist in the inhalation of eggs with dust.

The eggs pass in the faeces of definitive hosts. After ingestion by the intermediate hosts, the eggs hatch in the small intestine and releases an onchosphere, that penetrates the intestinal wall, then migrates through circulatory system into various organs such as liver and lungs. In these organs the onchosphere develops into a cyst, which grow gradually and producing protoscoleces. The definitive host become infected by ingesting the infected organs. After ingestion the protoscoleces evaginate, then attach to the intestinal mucosa and develop into adult

worm.

Adul in small intestine Protoscolex Irom cyst ingestion of cysts (m organs) Defusitive Host (dags & piner candae) Intermediate Host Ingestion of eggs (in fedes) isheep goats swine etc∤ egg in least 🛕 = Infective Stage Oncosphere natches 🛕 = Diagnostic Stage: penetrates intestural wall Hydaba cystinitives, lungs leto

Figure 1: Diagrammatic representation the life cycle of *E. granulosus*Indicate sheep-dog life cycle (Source: Parasites and health
Echinococcosis, Last modified: 2003).

The major differences in the life cycle of the four *Echinococcus* species are the nature of hosts involving (Rausch, 1997). The adult worm tend to host-specific, whereas the metacestode larva stages are able to develop in large number of different mammalian hosts-species. The geographic distribution of four species is important to the differentiate between the four *Echinococcus* species; *E. vogeli* and *E. oligarthus* are restricted to central and south America (Krammerer and Schantz, 1993). *E.multilocularis* distributed throughout the northern hemisphere extending from parts of central Europe into Asia and western Alaska, but recently occur in central and north America (Rausch, 1997). *E. granulosus* is the most widely distribution throughout the world in particularly common sheep herding areas of developing countries. In Libya, *E. granulosus* is the only species known to occur and responsible for hydatid disease in both human and domestic animals.

2.3 Strains of E, granulosus:

The term strain refers in the case of *E. granulosus* to « variants which differ statistically from other groups of the species in gene frequencies and in more characters of the actual of potential of hydatid disease » (Thompson and Lymbery, 1990; Thompson and McManus, 2002). For many years morphological and biological variability has been noted between populations of *E. granulosus* in different geographical regions and in different hosts. Described differences have included morphology, biochemistry, physiology, pathogenicity, development patterns and invectivity to human and domestic animals (Thompson and Lymbery, 1988).

The nature and diversity of variation within the genus *Echinococcus* have evolved rapidly in recent years through characterization of the

nuclear and mitochondrial genomes of representative isolates of the strains (Bowles et al., 1992; Bowles and Mc Manus, 1993a). At the present time, 10 genetically distinct population (genotypes) of E.granulosus from different geographical areas are known to exist (Lavikainen et al., 2003). The globally distributed G1 genotype of sheep is maintained in life cycles involving dogs and sheep. Its found in Australian mainland, Europe, USA, Newzealand, Africa, China, Middle East, South America and Russian Federation. The strain predominates in most domestic hosts in these countries (EcKert and Thompson, 1988 and Euzéby, 1991). Tasmanian sheep strain G2 genotype distributed in Tasmania and Argentine, its slightly different from the common sheep strain in mitochondrial DNA sequences . Buffalo strain G3 genotype in India. Horse strain G4 genotype, is adapted to horses and dogs throughout the united kingdom, Middle East, South Africa and Newzealand (Smyth and Davis , 1974 and Bowles et al., 1992). Cattle strain G5 genotype occurs in cattle and dogs in Europe, South Africa, India, Srilanka, Russian Federation and South America. Camel strain G6 genotype, in Middle East, Africa, China and Argentina in cycles involving camels and dogs. This strain was identified in Kenya, occurring in camels (Bowles et al., 1992 and Bowles and Mc Manus, 1993b). Pig strain G7 genotype, adapted to pigs and dogs has been reported in Europe, Russian Federation and South America (Mc Manus and Bryant, 1995). Cervid strain G8 genotype in North America and Europe, its cycles involving wolves and dogs. Lion strain G9 genotype in Africa (Thompson and Mc Manus, 2002). Fennoscandian cervid strain G10 genotype, in Finland (Lavikainen et al., 2003).

2.4 Biochemical profile of hydatid cysts:

McManus (1981) made a biochemical studies on the larval and adult stages of *E. granulosus*, he found that biochemical composition of *E. granulosus* of sheep origin in Kenya and United kingdom are identical and the constituents of DNA and RNA in both types were found similar in all the larval forms of *E. granulosus*. Sultan Sheriff *et al.* (1989) measured the levels of lipids and proteins in hydatid cysts fluid collected from lung and liver hydatid cysts of sheep and humans.

Shaafie et al. (1999) made a study on the biochemical profiles of hydatid cyst fluid of E. granulosus of human and animal origin from Libya. They found that there is quantitative variations in the levels of sodium, potassium, calcium, cholesterol, glucose, urea, creatinine and gama glutamyl transpeptidase in the cystic fluids of human and animal origins and reported that the similarities in the biochemical profiles of different hydatid cyst fluids suggest the existence of sheep strains of E. granulosus in human and other domestic animal in Libya. On the other hand Thompson and Lymbery (1991) suggested that biochemical studies on hydatid cysts from different host origins can provide valuable information to determine the existence of E. granulosus in Libya.

2.5 Epidemiology of hydatidosis:

The epidemiology of hydatid disease determined by the study of public health problem, risk factors, animal hosts and transmission cycles. Hippocrates described the human *E. granulosus* disease more than two thousand years ago with the peculiar term "liver filled with water". Al-Rahzes the famous Arabian physician, subsequently wrote on hydatid cyst of the liver about a thousand years ago (Adams, 1849). Hydatid disease still remains a serious health problem in certain parts of

the world and is endemic in Middle East, Mediterranean, South America and Australia (Erman et al., 2001).

Hydatid disease (E. granulosus) is endemic in many parts of the world. The annual incidence rates of human cystic echinococcosis in Northern Europe, Central Europe, Western Europe vary from sporadic, partially or mainly imported cases to low incidence rates of < 1 case per 100,000 inhabitants to higher rates up to > 10 cases per 100,000 (Navarrate et al., 1991; Löscher et al., 1992; Magnussen and Thorsen, 1992 and Braddick and Reily, 1993). Cystic echinococcosis has long been recognized as a serious public health problem in Greece (Karpathios et al., 1985), in Turkey (Altinatas and Ozensoy, 1993).

Echinococcus granulosus occurs at high rates of infection in most countries of the Middle East (Matossian et al., 1977). In endemic areas, the estimated range is 150 – 200 cases per 100,000 inhabitants (Gabriele et al., 1997). Important domestic intermediate hosts in the Middle East include the camels and sheep, with cattle, goats and donkeys also harbouring hydatid infections (Al-Abbassy et al., 1980; Al-Yaman et al., 1985: Abdel-Hafez et al., 1986a; Abdul-Salam and Farah, 1988 and Kamhawi and Hijjawi, 1992). The large stray dog populations found in Middle Eastern countries are an important reservoir hosts of the infection.

Information on the public health problems in Middle Eastern countries is largely based on reported clinical case series. In Jordan, the annual diagnostic incidence was estimated to vary from 15 to 65 cases per 100,000 in different regions of the country (Kamhawi and Hijjawi, 1992). In Saudi Arabia, numerous clinical reports suggest that a considerable public health problem may exist (Laajan and Nouh, 1991).

and Schaefer and Khan, 1991). In Iraq, hydatidosis is a disease of major public health importance, where over 500 human cases of the disease were reported annually (Matossian et al., 1977). In Iran, the average national surgical incidence of cystic hydatid disease varies between 0.1 (%) and 4.5(%) per 100,000 in different provinces (Mahdi and Benyan, 1990). An annual incidence is 3.6 (%) per 100,000 has been reported in Kuwaiti (Shweiki et al., 1990) but only 30% of the patients were Kuwaiti. In Syria, 157 patients, accounting for 2 % of all admissions at the University Hospital, were seen over a 7 years period (Munzer, 1991).

Christians than Muslims in Lebanon (Schwabe and Abou Daoud, 1961), and the disease was unknown in Somali Muslims in Kenya (Macpherson et al., 1989). These differences have often been assumed to be the result of differences in degrees of contact with dogs. Islamic religious believers consider the dog is an unclean animal and dogs are rarely allowed inside Muslim houses and human have little direct contact with dogs. These cultural differences may limit exposure of Muslim populations to dog-transmitted pathogen in comparison to their non-Muslim neighbours, they certainly have not excluded exposure to infection (Schantz et al., 1995).

Camels in east African region are an important intermediate host. However, E. granulosus occurring in this host as a distinct strain and rarely of ever infect humans (Schantz et al., 1995). Sheep and goats are the main intermediate hosts for sheep strain E. granulosus in Maasailand in Kenya (Macpherson, 1985) and probably in Ethiopia (Wosene, 1991). Goats and camels are thought to be the main host in Turkana, Kenya (Macpherson, 1981).

Most pastoral peoples keep large numbers of dogs where infection with *E. granulosus* have be reported in 3 – 87 % of dogs in Sudan (Saad and Magzoub, 1986), 39 – 70 % of dogs in Turkana, Kenya (Macpherson *et al.*, 1989) and 23 % in stray dogs in Somalia (Macchioni *et al.*, 1985).

In North Africa *E. granulosus* is propagated primary by a domestic cycle involving the domestic dog as the definitive host and many species of livestock including camels, cattle, sheep, goats; pigs, horses, donkeys and buffaloes as intermediate hosts (El-Kordy, 1946; Dar and Taguri, 1978; Larbaoui *et al.*, 1980; Jaiem, 1984; Gusbi *et al.*, 1987; Pandey *et al.*, 1988 and Ahmed, 1991). The local importance of each intermediate host species in maintaining the life cycle varies in different region, but this factor is often difficult to determine because many different domestic species infected. In all countries where the camel has been reported as an intermediate host, it is considered to be an important host for the local life cycle of the parasite.

The infection of dogs with E. granulosus have been reported from different countries, 20 % in Algeria (Senevet, 1951), 20 – 51 % in Morocco (Sirol and Lefevre, 1971; Pandey et al., 1987, 1988), 22 – 43 % in Tunisia (Jaiem, 1984; Behir et al., 1991), 8-38 % in Libya (Packer and Ali, 1986; Gusbi, 1987 and Awan et al., 1990) and 1 to 10 % in Egypt (Abo-Shady, 1980 and Hegazi et al., 1986). There are no dog control programmes in North Africa and the majority of dogs population have little contact with human (Gusbi, 1987; Shambesh et al., 1992).

In Tunisia between 800 and 1,200 new human cases of hydatidosis were reported every year for an annual incidence of 16.5 (%) per 100,000 (Gharbi et al., 1985 and Achour et al., 1989 and Anon, 1993). Hydatidosis is highly prevalent in Morocco, the annual incidence is 6.5 (%) – 7.8 (%) per 100,000 have been reported (Pandey et al., 1988; Ouhelli and Dakkak, 1992 and Kachani et al., 1998). Whilst in Algeria, 5,305 cases of hydatid disease were treated over a ten years period, giving an annual incidence of between 3.4 (%) to 4.6 (%) per 100,000 inhabitants (Larbaoui and Allyulya, 1979).

Human hydatidosis have been reported in Egypt by many authors (Babars et al., 1987; Romia et al., 1992; Mazyad et al., 1998 and Abu-Eisha, 1999). Haridy et al. (1998) among camels imported from Sudan for human consumption over five years reported 5.5 % (1992), 6.1 % (1993), 6.7 % (1994), 8.4 % (1995) and 4.3 % (1996) of camels were infected with hydatid disease.

2.6 Hydatidosis in Libya:

2.6.1 Human hydatidosis:

The prevalence of human hydatidosis depends on the degree of exposure of human to the source of infection, the strain of E, granulosus and individual resistance to infection. On the other hand both male and female of different age groups are equally susceptible to the infection with hydatid disease.

The first Libyan case of human hydatidosis in Benghazi was reported by Castigliola (1918). In 1927 Onorato detected 11 human cases during radiological survey in Tripoli Fossati (1970) estimated an incidence of 0.35 % in Benghazi (147 cases out of a population of 42000)

people) by using radiology these cases were confirmed by surgery . Between 1971 and 1976, about 0.70 - 0.85 % of the surgical cases, out of all admission, in Benghazi hospitals were infected with hydatid cysts (Dar and Taguri, 1978 and Kalani et al., 1984).

Dar and Taguri (1979) estimated that more than 20 % of hydatid infection was discovered accidentally. They mention that children and young adults under the age of 20 years constitute 20 – 25 % of all infected cases. El-Boulaqi and Taguri (1980) reported that of the 180 surgically confirmed hydatid cases recorded, 97 (53.9 %) were females and 83 (46.1 %) males from all over Eastern Libya. The most infected organ was liver (53.3 %), followed by lungs (30.6 %), Abdomen (7.8 %) and kidneys, the most susceptible age was below 24 years. Papadopoulos (1981) recorded age and sex distributions of 104 cases of the hydatid disease (operated surgically in chest hospital of Tripoli during the years 1972 – 1979). Rahman (1982) reported that 14 women had been found to be infected with pelvic hydatid disease in Benghazi during 1971 – 1979.

Gebreel et al (1983) indicated that a good number of Echinococcus infections were found among 200 inhabitants in El-Abiar with prevalence rate of 8 % in the urban area and 12 % in the surrounding rural area by using enzyme-linked immunosorbent (ELISA). Aboudaya (1985a) reported that 111 persons had been infected with hydatid disease among the 22979 patients admitted for surgery in Tripoli central Hospital a period of 1972 – 1979 with an overall incidence of 0.48 % of patients were females and 38 % males. The sites of infection were 88 liver infection (77.6 % of cases) followed by the muscle (4.4 %) abdomen (2.7 %) and lungs (1.7 %).

In 1992 Shambesh et al, carried out ultrasound study in five area of northwestern region of Libya put the prevalence at 2 %. Saad and Fatehell-Bab (1996) reported that 10 cases of hydatid disease were recorded in the Sebha University Hospital. The incidence of the hydatid disease within Libyan population is said to be 1 / 100 000 individual (Shambesh, 1997).

In 1999, the result of a large-scale nationwide survey of cystic echinococcosis, based on ultrasound examinations, revealed that those are living in the northern coastal area had the disease (Shambesh et al., 1999). Khan and El-Buni (1999) collected the finger-prick blood samples on filter paper discs for the presence of anti-hydatid antibodies from University students in Benghazi where the study revealed that sero-positive rate was 1.66 %. Al-Saqur et al (2001) reported that 1.03 % of humans were infected with hydatidosis in Southern Libya mainly in Sebha. Tashani et al (2002) recorded that 306 surgically confirmed human cases of cystic echinococcosis were identified in the medical records studied in Benghazi and reported that the post-operative reports for 215 of the patients which indicated that most hydatid cysts were found in liver (65 %), followed by the lungs (13 %), spleen (5 %), bones (2 %) and kidneys (2 %).

Mohamed et al. (2004) reported that the prevalence in humans from Sebha Hospital during the years 2002 and 2003 was 1.103 % from the total cases of admitted in the Surgery Department, they mention that, this not necessarily reflect an actural incidence rate in Sebha.

The scrological techniques were used in some studies to determine the prevalence of human hydatidosis (Gebreel et al., 1983; Khan et al., 1990; Shambesh et al., 1992; 1999 and Khan and Kidwai, 1996). These studies indicate the infection rates but do not clarify the relationship between the presence of antibodies and clinical signs and current presence of hydatid cysts.

2.6.2 Livestock hydatidosis:

The earliest known reports on infection rates in intermediate hosts are those of Medulla (1931) and Cicogna (1961) reported a prevalence rate of 40 % in sheep and 70 % in cattle in slaughterhouse in Tripoli were infected with hydatid disease. Gebreel *et al.* (1983) reported the incidence (%) of the disease in herbivorous animals in slaughterhouse of Benghazi during the years 1975 – 1977.

Aboudaya (1985b) collected statistics on hydatid cysts in domestic animals from 46 slaughterhouses distributed throughout Libya. The rate of infection in Libyan reared animals was highest in camels (27 %) and lowest in sheep (4.3 %), with a low to moderate infection rate of 6.6 % in cattle. The study carried out by Gusbi *et al* (1987) on the incidence and geographical distribution of hydatid cysts in sheep from ten localities in Libya revealed that 12.74% of adult sheep and 0.29 % of lambs were found infected with hydatid cysts. The most infected organ was liver (97.26 %) followed by lungs (58.70 %), kidneys (1.79 %), spleen (0.74%) and heart mesentery and muscles (0.24 % each). They suggested that the high cyst prevalence rate and high fertility in sheep indicates that the sheep / dog « strain » of *E. granulosus* is the most widespread strain in Libya.

Gusbi et al. (1990) in Kufra reported that 50 % of hydatid cysts infection in camels which reflects the preciousness of these animals to

Prevalence rates in goats and cattle were 1.5 % and 4.5 % respectively. The total fertility of goat cysts was 61.7 % and most of these cysts occurred in lungs. All cattle cysts were completely sterile (Gusbi $et\ al\ ,$ 1990). In 1997 Ibrahem and Gusbi described camels as maintenance hosts in all north country—with sheep , goats and cattle playing a secondary role in the Life cycle of E, granulosus.

The study conducted by Ibrahem and Craig (1998) on slaughtered animals in different areas of northern Libya, they found that the prevalence rates of hydatidosis in sheep, goats and camels were 15.8%, 3.8% and 48%. About the site of infection, the liver was predominant infected site with prevalence rates of 86% in sheep, 100% in goats, while in camels, lung was the most infected organ (85.4%) and liver (33%), they indicated that more than 90% of the camel hydatid cysts were fertile.

Gdourra (2003) made a study on 1380 local camels slaughtered at the abattoirs in Sirt city. The prevalence of hydatid cysts was 3.62 %. The results showed that the infection was not age dependent and differences in prevalence between male and female camels were statistically insignificant.

Mohamed et al. (2004) conducted a study on 1022 camels . 200 sheep , 85 cattle and 50 goats slaughtered at the abattoirs in Sebha city . Those animals were inspected for hydatid cysts of E . granulosus, the prevalence of hydatidosis was 12.62 % in camels , 18.5 % in sheep and 4% in goats, and there were no differences in prevalence between male

and female camels. However, older camels had more infection than younger one.

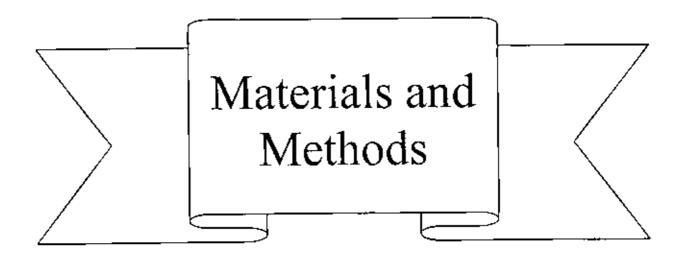
2.6.3 Definitive host Hydatidosis:

The dog population in Libya can be divided into three groups: (1) stray dogs are found in all urban and rural areas, (2) farm dogs are those kept by farmers to guard the animals or farm property, (3) shepherd dogs and (4) pet or house dogs. The scarcity of data of adult worm E, granulosus in dogs may due to the fear of infection discourages the examination of dogs for the determination of prevalence of the adult parasite. The first study on the prevalence of the disease in dogs was recorded by Cicogna in 1961, were found that 60 % in the shepherd dogs and 10% in the town dogs were infected with E, granulosus. Packer and Ali (1986) conducted a survey on dogs in Tripoli, they found that the prevalence rate of 11.8 % of stray dogs and 5.9 % of farmer dogs, while no E, granulosus infection was recorded from pet dogs.

In 1987 Gusbi, carried out a survey in 14 localities in Libya, the overall prevalence rate reported was 27.81 %, the infection rates reported for stray, sheep, domestic and semi domestic dogs were 40.3 %, 34.8%, 7.7 % and 9.3 % respectively. The study indicated that the incidence rates in dogs in southwestern region of Libya are quite low, 6.7 % in Sebha, 0 % in Gdames, Socna and Ghat, while the prevalence of the worms was generally higher in the whole coastal areas. The same study revealed that 80 % of dogs were infected in Tobruk, which is markedly high rate. Awan et al., 1990, conducted a survey on the prevalence of Echinococcus infection in dogs from five Libyan cities, and found that 36.8 % and 35.3 % of stray and semi domesticated dogs were infected, and the load of infection varied between 1 to 12821 worms per dog

El-Sageyer (1989) reported that the incidence rates in dogs in Benghazi and Tripoli were 7.14 % and 26.19 % respectively.

Generally, in Libya, the majority of the dog population have little contact with human (Gusbi, 1987, Shambesh et al, 1992). The dogs are kept to guard sheep and goats from predatory stray dogs and wolves and usually more than one dog is found per household (Shambesh et al, 1992).



3. MATERIAL AND METHODS

3.1. Study area:

Sirt is located on the Mediterranean coastal some 450 km east of Capital Tripoli (Fig. 2). It occupies an area of 69 km² most of it is semiarid area, with population of about 140,000 (Source: National information Board, Sirt 2001). The warmest months of the year are July and August where temperature may rise to 42°C, while the coolest months are January and February where temperature may decline up to 7°C. The relative humidity ranges from 67 to 71 % throughout the year. The average rainfall exceeds 253 mm. (Source: Sirt Meterological Office, Personal Communication 2001).

3.2. Animals:

There are about 1 470 000 sheep and 950 000 goats in Sirt, the later are raised on state owned farm within the city. There are about 110 000 camels in the Sirt areas (Sirt agriculture and animals resource office, 2005).

3.2.1. Studied animals:

The animals which slaughtered at Sirt abattoir used in this study including: sheep (Ovis aries) the famous Libyan Barbary breed type. There are characterized by their coarse wool, fat tail and straight or slightly convex nasal profile (Devendra and Mcleroy, 1982) (plate 1a and 2), goats (Capra hircus) (plate 2) and camels (Camelus dromedaries) both considered by abattoir veterinarians to be indigenous species and therefore of native Libyan breeds (plate 3). Cattle (Bos Taurus) breeds studied were of three types, Friesian, Jersey and local (plate 4a and b).

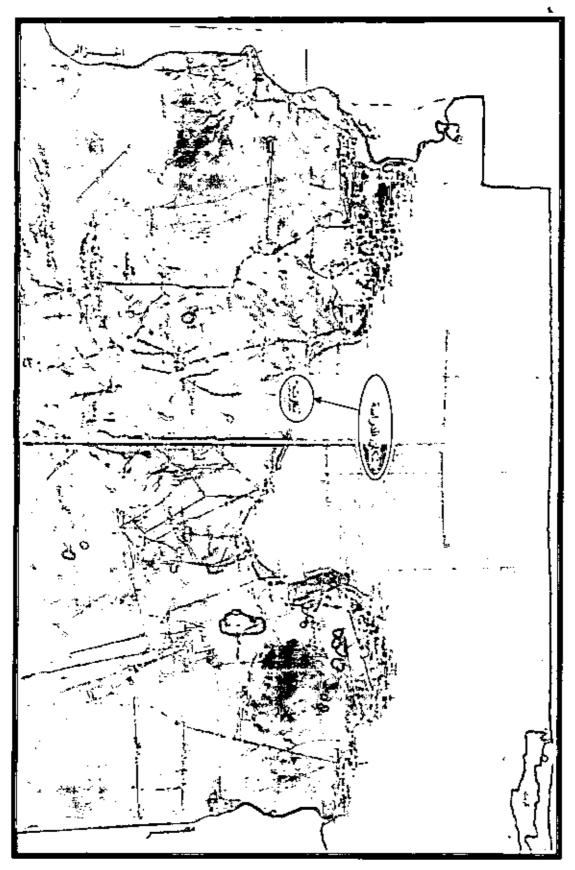
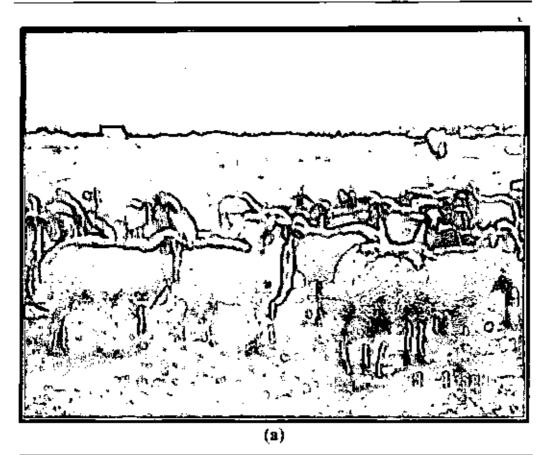


Figure 2: Map of Libya-showing place of the study (Sirt city).



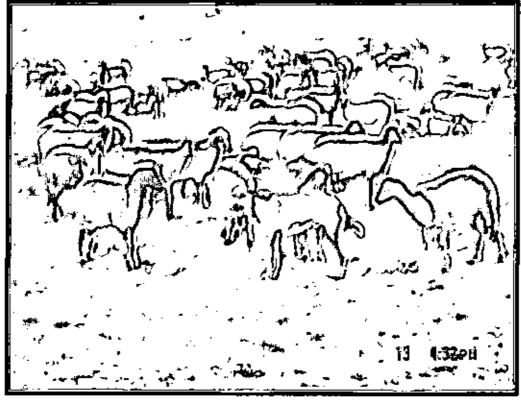


Plate 1a, b: Libyan Bar bary sheep (Ovis aries).

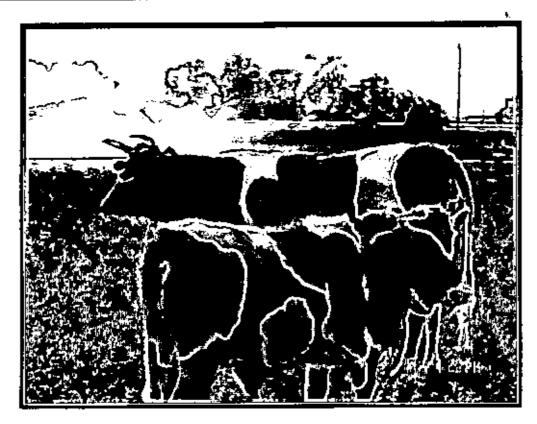
(b)



Plate 2 : Goats (Capra hircus) .



Plate 3: Libyan camels (Camelus dromedaries).



(a)

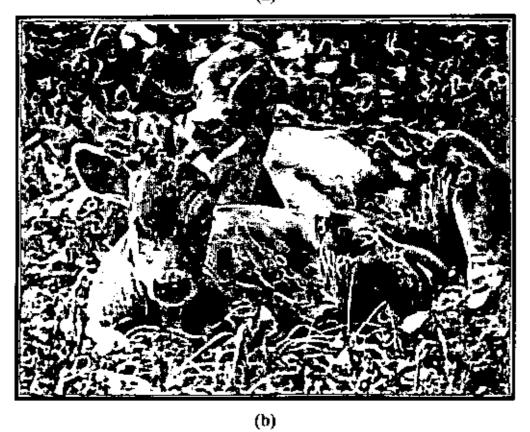


Plate 4a, b: Cattle (Bos taurus) bread Friesian (a), local (b).

The age of these animals were determined by conventional dentition method (Ensminger, 1983).

3.3. The abattoirs:

Sirt city has central authorized abattoir. It is housed in new buildings in which the slaughtering and processing of carcasses are carried out manually (plate 5 and 6). All animals included in the present study were slaughtered at the central abattoirs during the period from July 2004 to Jun 2005. The abattoirs was visited twice a week.

3.4 Detection of parasite:

Over a year from July 2004 to Jun 2005, a total of 12769 animals (3794 sheep, 8123 goats 739 camels and 113 cattle) were examined for hydatid cysts. The abattoir was visited early in the morning in order to attend the initial segregation of slaughtered animals, during each visit, sex, age and breed of each animal was recorded. Liver and lungs were fully and carefully examined for the presence of hydatid cysts (plate 7), in addition heart, kidneys and spleen were examined in cases of heavy infections. Criteria used for the identification of cysts were those described by Gracey and Collins (1992).

3.5 Examination of hydatid cysts:

Liver and lungs were the organs usually examined for hydatid cysts. All detected cysts were incised and subjected to macroscopic examination. Infected organs were carefully washed with water to remove debris and blood, then placed separately in labeled plastic bags and transported to the laboratory of Zoology Department Faculty of Science, Al-Tahadi. University as soon as possible for examination. The total number of cysts

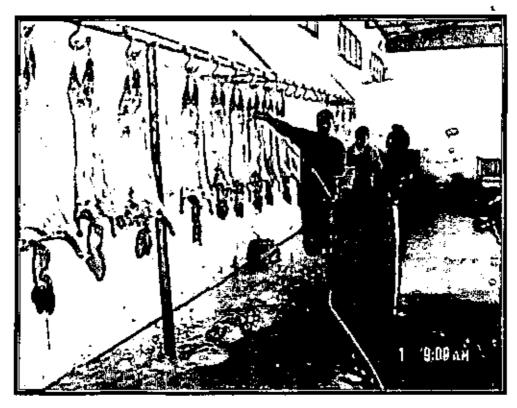


Plate 5: Sheep hall of Sirt abattoir showing manual processing of carcasses.



Plate 6 : Slaughtered sheep investigated by abattoir veterinarian to find out any hydatid cysts .



Plate 7: Inspection of sheep liver for hydatid cysts.

for each infected organ was recorded (plate 8), and five cysts (if possible) from each organ were selected for further examination.

3.6 Determination of hydatid cyst characteristics:

3.6.1 Cyst size: The cyst size (length and width) was measured by ruler in centimeters and recorded:

Hydatid cyst fluid was aseptically aspirated using sterile syringes within few hours after collection of hydatid cysts, and the fluid was emptied into graduated cylinder and the hydatid fluid volume was determined. A part of hydatid fluid was emptied into glass petri dishes, the cysts were opened and carefully examined for the presence of daughter cysts, hydatid sand and germinal layer. The germinal layer was removed and taped onto the hydatid fluid to release broad capsules.

3.6.2 Estimation of hydatid cyst fertility:

A drop of hydatid cyst fluid was placed onto a glass slide, then examined by light microscope for the presence of broad capsules or protoscoleces. The cyst considered as fertile when its fluid contained protoscoleces, while those without protoscoleces considered as sterile cysts (plate 9).

3.6.3 Estimation of hydatid cyst viability:

The viability of hydatid cyst was tested (1) using 0.1 % eosin stain test (Himonas et al., 1994), this test show that those protoscoleces stained with eosin stain were considered as dead or non-viable protoscoleces, while the viable protoscoleces did not stained and appeared transparent (plate 10). (2) published morphological descriptions of hydatid cyst protoscoleces (Saidi, 1976 and Thompson, 1995). This



Plate 8: Examination of livers and collection of hydatid cysts.



Plate 9: Microscopic examination fof hydatid cysts and studding the morphology of protoscoleces and rostellum hooks in fertile cysts.

test depends on the colour , size , and their muscular contraction of protoscoleces . Cyst viability was calculated as the percentage of viable protoscoleces . This was by using aliquots of 100 μL of hydatid fluid containing hydatid sand , for each cyst examined spread on microscope slide , then examined under a light microscope (plate 11) .

3.6.4 Morphological studies on rostellar hooks:

Fertile cysts of liver and lungs of sheep, goats and camels were used in the study of morphological features of rostellar hooks (Table 1). 25 cysts were randomly selected from each organ and 8 protoscoleces were measured for each cyst. 25 large and 25 small hooks used for examination were recorded and measured for each protoscolex. Viable protoscoleces were used and their viability was determined using the method described above. The arrangement of hooks and their total number in each protoscolex was recorded. The measurements of the total hook length, blade length, handle length and width for large and small hooks were done according to the method described by Sweatman and Williams (1963) (plate 12). This was studied by using a calibrated Carl Ziess microscope at a magnification of x 100 (Oil immersion). Protoscoleces and hooks were photographed using a Carl Ziess photomicroscope.

3.7 Statistical analysis:

A computerized statistical program SPSS. V11.5 (Statistical Package Science Sociality) has been used to analyze the data of the present study. Prevalence was calculated as the percentage of infected animals and the mean intensity as the mean number of cysts per infected animal. The significance levels were determined by using Pearson and Spearman rank

correlation . The accepted level of significance was $P \leq 0.05$ and stong significance was $P \leq 0.01$.

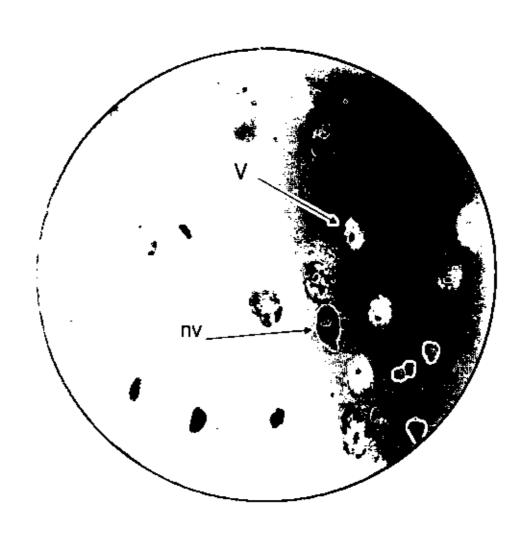


Plate 10: Hydatid cysts showing viable (v) and non-viable (nv) protoscoleces following treatment with 0.1 % eosin to test viability (x = 10).

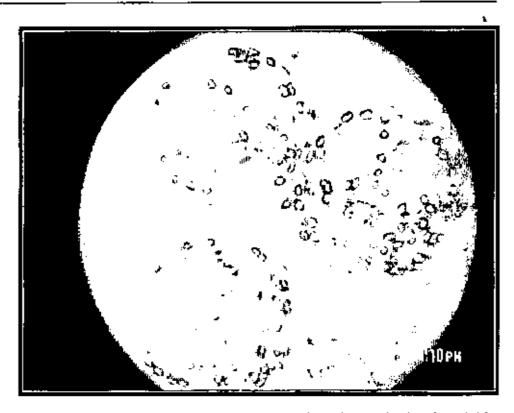


Plate 11 : Hydatid cyst from sheep liver , broad capsule showing viable and non-viable protoscoleces (X=40).

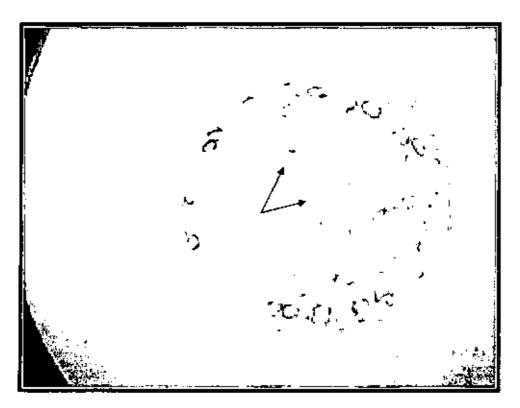
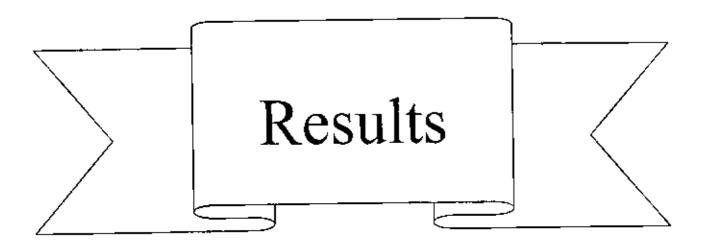


Plate 12: Protoscoleces from sheep liver hydatid cysts showing the arrangement of rostelum hooks (X = 100).

Table 1: Number of rostellum hooks from liver and lung hydatid cysts of Echinococcus granulosus examined per host species.

												Ī
Animal species (N)		Sheep			Goats	·		Camels	:		Cattle	
Organ	Liver	Liver Lungs	Total	Liver	Lungs	Total	l'iver	Lungs	Total	Liver	Lungs	Total
No. of organs	10	01	20	13	<u> </u>	20	14	16	20	18	12	30
No. of cysts	25	25	50	25	25	50	25	25	50	25	25	50
No. of protoscoleces	100	100	200	100	100	200	100	001	200	0	0	0
Total no. of hooks	001	100	200	100	001	200	100	100	200	C	0	0



4. RESULTS

Hydatidosis has been reported from different communities where dogs, the main definitive host, have been access to raw offal through home-slaughter from poorly regulated abattoirs or from scavenging carcasses or discarded offal (plates 13, 14, 15 and 16).

The annual slaughter rate in Sirt was determined by the total number of animals slaughtered at Sirt abattoir. During the period from July 2004 to June 2005, 13136 sheep, 30099 goats, 239 cattle and 3233 camels were slaughtered at Sirt abattoir.

4.1. Prevalence:

The hydatid cysts of *Echinococcus granulosus* were detected in all animal species examined. A total of 186 (4.9 %) of 3794 sheep, 195 (2.4%) of 8123 goats, 17 (15 %) of 113 cattle and 20 (2.7 %) of 739 camels were found to be infected with hydatid cysts (Table 2 and Fig. 3).

4.1.1 Prevalence according to sex:

The prevalence of hydatidosis in both males and females of intermediate hosts (sheep, goats, cattle and camels) shown in Table (3) and Fig. (4). The results have shown that prevalence in males sheep was 5.9%(73/1247) and females was 4.4%(113/2547). No significant differences in prevalence of hydatidosis between both sexes (P = 0.58). Prevalence of hydatidosis in female goats 3.9%(90/2287) was higher than that detected in males 1.8%(105/5836). The results show significant differences in prevalence between males and females of goats (P = 0.000). While the prevalence in males and females cattle was 12.9% (9/70) and 18.6% (8/43) respectively, the results show no significant



Plate 13: The method by which the proper disposal of condemned organs.

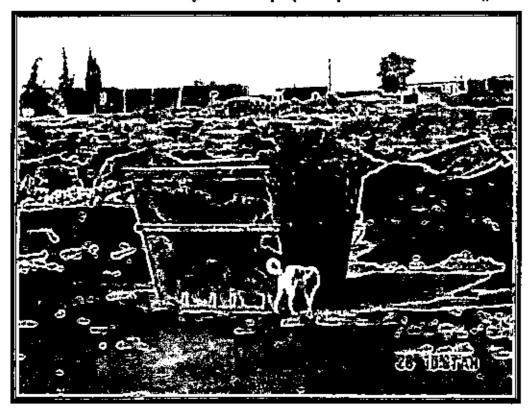


Plate 14: Dogs (The main definitive host) near the place where the disposal of condemned organs.

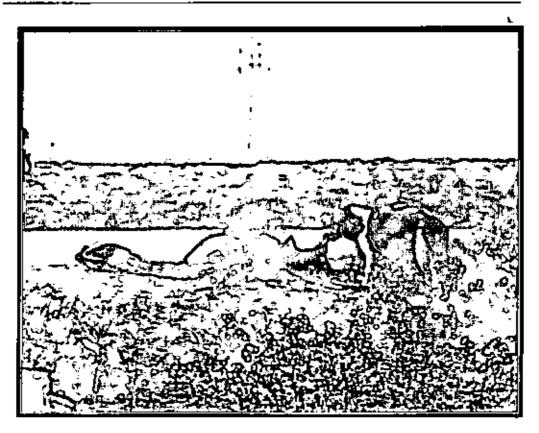


Plate 15: Showing the dog-definitive host feeding on dead camel.

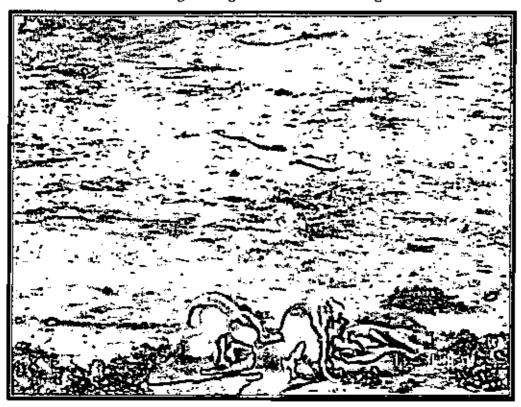
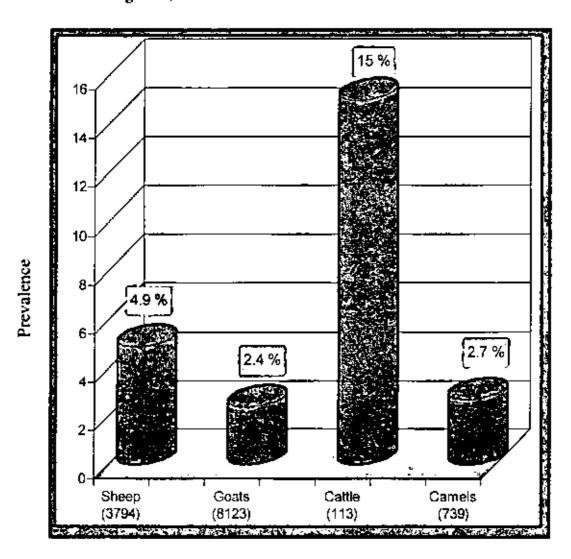


Plate 16: Sirt dump-Note the dogs feeding on disposal of condemned organs .

Table 2: Overall prevalence of hydatid cysts in slaughtered animals hosts in Sirt.

Examined animals	No. examined	No. of infected (%)
Sheep	3794	186 (4.9)
Goats	8123	195 (2.4)
Cattle	113	17 (15.0)
Camels	739	20 (2.7)

Figure 3: Overall prevalence of hydatid cysts in slaughtered sheep, goats, cattle and camels in Sirt.



Examined animals

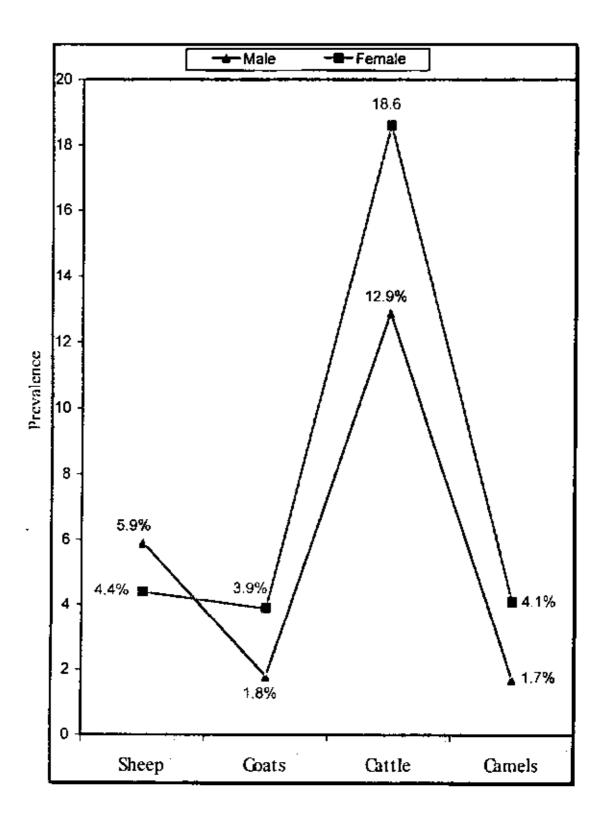
Table 3: The prevalence of hydatid cysts in slaughtered animals according to their sex.

Sex	S	Sheep)	Goats	5	Cattle	Ü	Camels
	No. examined	No. infected (%)	No. cxamined	No. infected (%)	No. examined	No. Infected (%)	No. examined	No. infected (%)
Male	1247	73 (5.9)	9836	105 (1.8)	70	9 (12.9)	421	7(1.7)
Female	2547	113 (4.4)	2287	90 (3.9)	43	8 (18.6)	318	13 (4.1)
Total	3794	186 (4.9)	8123	195 (2.4)	113	17 (15.0)	739	20 (2.7)
p-value	D :	0.058	D	0.000	0.411	0.411 (Non-sig)	0.	0.044

*** Correlation is significant at the 0.01 level (2-tailed).

^{*} Correlation is significant at the 0.05 level (2-tailed).

Figure 4: Prevalence of hydatid cysts in slaughtered animals according to their sex.



differences between both sexes (P = 0.411). The prevalence of hydatid cysts in males of camels was 1.7 % (7/421) while in females was 4.1 % (13/318). There was a slight correlation and significant difference (P = 0.044) in the prevalence of hydatidosis between both sexes.

4.1.2 Prevalence according to age:

The relationship between prevalence of hydatidosis and different age groups shown in (Table 4). The results have shown that infection is age dependent in sheep , the older animals had higher the prevalence. The infection rates were 3.0 %, 4.6 %, 7.1% and 12.6 % detected in age groups < 1, 1 - < 2, 2 - < 3 and ≥ 3 years respectively (Fig. 5). The results have shown strong correlation and significance between age groups of sheep (P = 0.000). The same trend was seen in goats with the older group (goats older than three years) having a prevalence of 10 % which contrasts with less than 1.5% in goats younger than 1 year (Fig. 6). The results shows a strong correlation and significant difference between age groups of goats (P = 0.000).

Prevalence of infection in cattle older than 3 years was 22.7 % This was significantly higher than 2.1 % seen in cattle younger than 1 year and the 14.3 % in the age groups ranging between 1-2 years (Fig. 7). Among camels, infection with age was similar to that of the other animals species. The infection rates were 0.8 %, 2.5 %, 7.3 % and 9.2 % detected among age groups <1, 1-<2, 2-<3 and \geq 3 years (Fig. 8) The differences between age groups were statistically significant (P = 0.000).

4.1.3 Prevalence according to season:

Infection of hydatid cysts among all animal species were recorded throughout the year and the results showed that there is no correlation and a significance difference between the four seasons (winter, spring, summer and Autumn). The infection rates during the Winter, Spring, Summer and Autumn were 4.6 %, 4.8 %, 4.9 % and 5.4 % respectively among sheep, 3.2 %, 2.7 %, 1.8 % and 2.6 % respectively among goats, 15.4 %, 13.0 %, 12.5 % and 17.8 % respectively among cattle and 2.5%, 2.7%, 2.9% and 2.7% respectively among camels (Table 5 and Figs. 9, 10, 11, 12).

Table 4: The prevalence of hydatid cysts in slaughtered animals according to their age.

								·—
		Sheep		Goats		Cattle	ا ت 	Camels
Age (Years)	No. examined	No. infected (%)	No. examined		No. examined	No. infected (%)	No. examined	No. Infected (%)
< 1	1920	58 (3.0)	4062	61 (1.5)	47	1 (2.1)	393	3 (0.8)
1-<2	947	44 (4.6)	2422	40(1.7)	2]	3 (14.3)	199	5 (2.5)
2-<3	592	42 (7.1.)	1141	44 (3.9)	23	8 (34.8)	82	6 (7.3)
>3	3	42 (12.5)	498	50 (10.0)	22	\$ (22.7)	99	6 (9.2)
Total	3794	186 (4.9)	8123	195 (2.4)	113	17 (15.0)	739	20 (2.7)
p-value	0.	0.000***	0	***	0	0.001 **	0.0	0.000 ***
						!		

Correlation is significant at the 0.01 level (2-tailed).

Table 5: The prevalence of hydatid eysts in slaughtered animals according to their seasons.

	Š	Sheep	9	Goats		Cattle	Ca	Camels
scasons	No. examined	No. infected	No. examined	No. infected	No. examined	No. infected	No. examined	No. Infected
Winter	610	28 (4.6)	6811	38 (3.2)	13	2 (15.4)	163	(%) 4 (2.5)
Spring	1093	52 (4.8)	2112	56 (2.7)	23	3 (13.0)	146	4 (2.7)
Summer	1276	62 (4.9)	2845	50 (1.8)	32	4 (12.5)	207	6 (2.9)
Autumn	815	44 (5.4)	1977	51 (2.6)	45	8 (17.8)	223	6 (2.7)
Total	3794	186 (4.9)	8123	195 (2.4)	113	17(15.0)	739	20 (2.7)
p-value	0	0.476	0	0.160		0.686	0	0.895

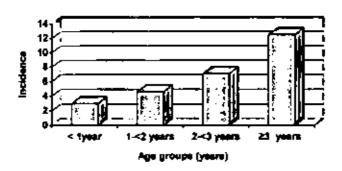


Figure 5: Prevalence of hydatidosis in sheep at different age groups

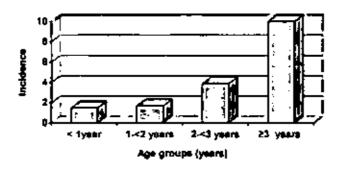


Figure 6: Prevalence of hydatidosis in goats at different age groups

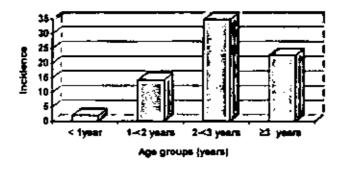


Figure 7: Prevalence of hydatidosis in cattle at different age groups

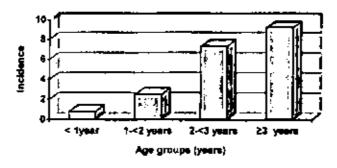


Figure 8: Prevalence of hydatidosis in camels at different age groups

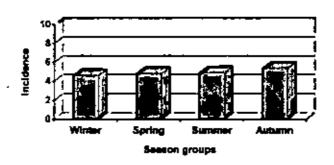


Figure 9: Prevalence of hydatidosis in sheep at different seasons

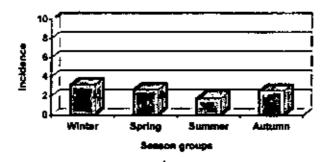


Figure 10: Prevalence of hydatidosis in goats at different seasons

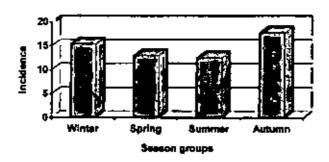


Figure 11: Prevalence of hydatidosis in cattle at different seasons

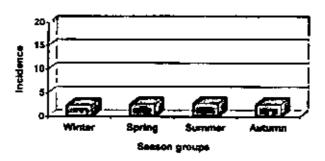


Figure 12: Prevalence of hydatidosis in camels at different seasons

4.2 Location of hydatid cysts:

The most commonly infected organs in all examined animal species were the liver and the lungs. Extra-hepatic and extra-pulmonary cyst locations (e.g. spleen, kidney, body cavity) constituted less than 0.5 % of all infections and have therefore excluded from the analysis. The prevalence of the hydatidosis in liver and lungs of sheep were 77.9 % and 48.9 % respectively.

Liver infection in goats was encountered in 73.2 % of all cases, but lungs were also infected with $60\,\%$. Cattle cysts were seen in the liver in $64.7\,\%$ of infected cases, whereas lung infection constituted $58\,\%$ of these infections. Lungs were the most infected organs in camels $75\,\%$ of infected cases and liver infection was $50\,\%$ of these cases.

On the other hand the results shown that infected liver, lungs and both liver and lungs in sheep were encountered in 51.1 %, 22.6 % and 26.3 % respectively. The infection in liver was higher than lungs and liver and lungs together. In goats liver infection was 39 %, lung infection was 27.7 % and infection in liver and lungs together was 33.3%. The results showed that liver infection in cattle was encountered in 41.2 % and lungs infection in 35.3 %, liver and lungs together constituted 23.5 %. Infection in camels showed that lungs infection 50 % was higher than liver infection 25 % and liver and lungs together 25 % (Table 6 and Fig. 13).

The results showed strong correlation and there is a significant differences between age groups of sheep and goats and location of cysts (P=0.000). The age had no effect on cyst location (Table 7). The sex of animals had no effect on cyst location of cysts in organs in sheep,

Table 6: Location of hydatid cysts in liver and lungs of sheep, goats, cattle and camels.

% of	infected o	rgans out of t	otal infected a	nimals
Examined animals	No. infected	Infected liver (%)	Infected lungs (%)	Infected liver&lungs (%)
Sheep	186	95 (51.1)	42 (22.6)	49 (26.3)
Goats	195	76 (39.0)	54 (27.7)	65 (33.3)
Cattle	_17_	7 (41.2)	6 (35.3)	4 (23.5)
Camels	20	5 (25.0)	10 (50.0)	5 (25.0)

Figure 13: Location of hydatid cysts in liver and lungs of sheep, goats, cattle and camels.

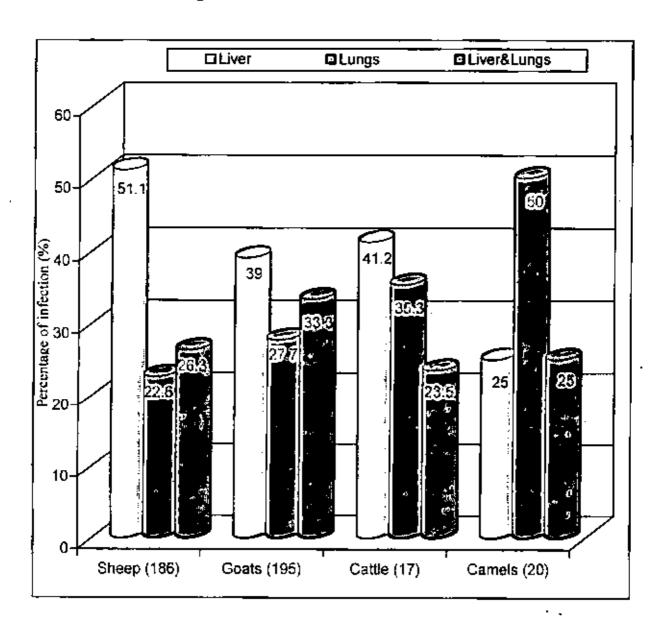


Table 7: Location of hydatid cysts in the liver and lungs of the sheep, goats, cattle and camels according to their

Age Infected Infected <th< th=""><th></th><th><u> </u></th><th>Sheep</th><th></th><th></th><th>Goats</th><th></th><th></th><th>Cattle</th><th></th><th></th><th>Camels</th><th>. . </th></th<>		<u> </u>	Sheep			Goats			Cattle			Camels	. .
Infected	05.4								1 - 6 - 6 - 6 - 6	Infortac	Infocted	Infected	Infected
liver lungs liver& liver& liver& liver 40 12 6 32 24 5 0 1 25 14 5 19 16 5 1 1 19 10 13 10 21 5 1 1 11 6 25 12 4 34 0 3 11 6 25 12 4 34 0 3 11 6 25 12 4 34 0 3 11 6 25 12 4 34 0 3 11 6 25 12 4 34 0 3	3#4	Infected	Infected	Infected	Infected	Infected	Infected		параши				
40 12 6 32 24 5 0 1 25 14 5 19 16 5 1 1 19 10 13 13 10 21 5 1 11 6 25 12 4 34 0 3 11 6 25 12 4 34 0 3 0.000*** 0.000*** 0.000(Non-signature)	(Years)	liver	lungs	liver&	liver	Jungs	liver&	liver	säuni	liver&	liver	kgunl	liver&
40 12 6 32 24 5 0 25 14 5 19 16 5 2 19 10 13 13 10 21 5 11 6 25 12 4 34 0 11 6 25 12 4 34 0 0.000**** 0.000*** 0.000*** 0.000*** 0.0000 0.000 0.0000 0.0000 0.0000 0.0000 0.0000 0.0000 0.0000 0.0000 0.0000 0.0000 0.0000 0.0000 0.0000 0.00000 0.0000 0.0000				lungs			lungs			Spant			lungs -
25 14 5 19 16 5 2 19 10 13 13 10 21 5 11 6 25 12 4 34 0 0.000*** 0.000***	 	40	12	9	32	24	S	0	-	0	0	3	0
25 14 5 19 16 5 2 19 10 13 10 21 5 11 6 25 12 4 34 0 11 6 25 12 4 34 0 0.000**** . 0.000 *** 	;	<u> </u>							†				,
19 10 13 13 10 21 5 11 6 25 12 4 34 0 0.000*** 0.000 ***	1-2	25	14	· S	61	16	'n	۲۱	-	0	0	_	o
19 10 13 13 10 21 5 11 6 25 12 4 34 0 0.000*** 0.000 ***								,		,	ŗ	٠ ،	r
11 6 25 12 4 34 0 0.000*** 0.000 ***	2-3	61	10	13	<u></u>	2	-5	<u>د</u>	_	7	7	`	,
0.000***	 - -				٤		2.1	٥	~	2	C	m	ری
*** 00000	× 3	=	9	25	71	4	÷.)	- -	 			
0.00.0						** 0000			160 (Non	-sir)	-	(gis-noN) 00.1	(<u>e</u>
	p-value		0.000			00000						, , 	: -

Table 8: Location of hydatid cysts in the liver and lungs of the sheep, goats, cattle and camels according to their sex.

											-	
	j	Sheep			Goats			Cattle			Camels	
Sex	lafected liver (%)	Infected Infected Infected liver Lungs liver&		Infected liver (%)	Infected lungs (%)	Infected liver& lungs (%)	Infected liver (%)	Infected Lungs (%)	Infected liver& lungs (%)	Infected liver (%)	Infected lungs (%)	Infected liver& lungs (%)
Males	36 (49.3)	16 (21.9)	<u> </u>	49 (46.6)	_28 (26.7)	28 (26.7)	4 (44.4)	4 (44.4)	(11.1)	(28.6)	(57.1)	(14.3)
Females (52.2)	59 (52.2)	26 (23.0)	28 (24.8)	27 (30.0)	26 (28.9)	37	(37.5)	2 (25.0)	(37.5)	(23.1)	(46.2)	(30.7)
Total	95	42	49	78	54	9		9	4	. 2	10	2
p-value	0.5	0.589 (Non-sig)	sig)		0.011 *		0.4	0.414 (Non-sig)	sig)	0.5	0.533 (Non-sig)	sig)
			į			İ		ı				

cattle and camels. While there is a significant difference between sex of goats and cyst location (P = 0.01) (Table 8).

4.3 Intensity of infection:

The number of cysts in sheep varied from 1 to 20 cysts in both liver and lungs , 53.8 % (1-3 cysts) , 36.0 % (4-8 cysts) and 10.2 % (9-20 cysts) . Goats showed 61.6 % (1-3 cysts) , 28.7 % (4-8 cysts) and 9.7 % (9-20 cysts) . Cattle showed 52.9 % (1-3 cysts) , 35.3 % (4-8 cysts) and 11.8 % (9-20 cysts) . Camels showed 40 % (1-3 cysts) , 35 % (4-8 cysts) and 25 % (9-20 cysts) . (Table 9 and Fig. 14) .

4.3.1 Intensity of infection and age:

The results showed that a significant difference between age groups and intensity of infection in sheep and goats (P=0.00). The intensity of infection in organs of sheep and goats increase with age increase. On the other hand there was no correlation and a significant difference between age and intensity of infection in cattle (P=0.699) and camels (P=0.053) (Table 10).

4.3.2 Intensity of infection and sex:

The results showed that there is no correlation between the intensity of infection and sex of sheep, goats, cattle and camels (Table 11).

Table 9: Intensity of infection in liver and lungs of sheep, goats, cattle and camels.

-	Intensity o	f infection	
Examined animals	1 – 3 cysts (%)	4 – 8 cysts (%)	9 – 20 cysts (%)
Sheep (186)	100 (58.8)	67 (36.0)	19 (10.2)
Goats (195)	120 (61.6)	56 (28.7)	19 (9.7)
Cattle (17)	9 (52.9)	6 (35.3)	2(11.8)
Camels (20)	8 (40.0)	7 (35.0)	5 (25.0)

Figure 14: Intensity of infection in liver and lungs of sheep, goats, cattle and camels.

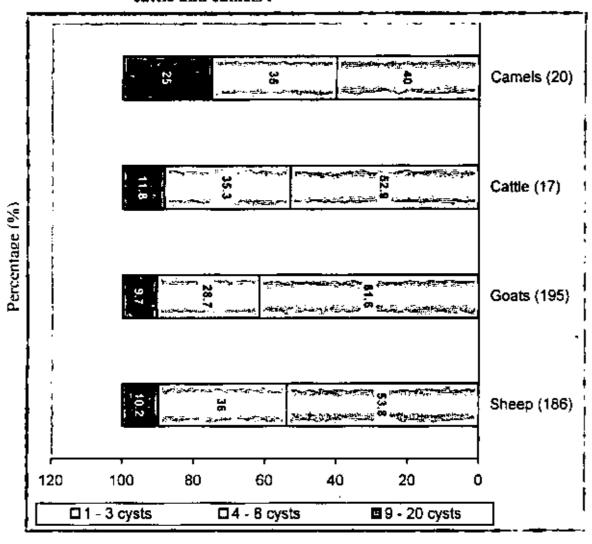


Table 10: Intensity of infection in sheep, goats, cattle and camels according to their age.

	_ 	Inter	nsity		
Examined animals	Age (Years)	1 – 3 cysts (%)	4 – 8 cysts (%)	9 – 20 cysts (%)	p-value
	<1	43 (74.1)	13 (22.4)	2 (3.5)	
Sheep	1-<2	26 (59.1)	18 (40.9)	0 (0.0)	0.000***
186	2-<3	18 (42.9)	18 (42.9)	6 (14.2)	
	<u>≥</u> 3	13 (31.0)	18 (42.9)	11(26.1)	
	<1	54 (88.5)	7 (11.5)	0 (0.0)	
Goats	1-<2	22 (55.0)	13 (32.5)	5 (12.5)	0.000***
195	2-<3	23 (52.2)	16 (36.4)	5 (11.4)	
ļ	≥3	21 (42.0)	20 (40.0)	9 (18.0)	
	<1	1 (100.0)	0 (0.0)	0 (0,0)	
Cattle 17	1-<2	1 (33.3)	1 (33.3)	1 (33.4)	0.699
	2-<3	6 (75.0)	1 (12.5)	1 (12.5)	
	≥3	1 (20.0)	4 (80.0)	0 (0.0%)	
	<1	3 (100.0)	0 (0.0)	0 (0.0)	
Camels	1-<2	0 (0.0)	1 (100.0)	0 (0.0)	0.053
20	2-<3	4 (44.4)	4 (44.4)	1 (11.2)	
į	≥3	1 (14.3)	2 (28.6)	4 (57.1)	! ! ! !

Table 11: Intensity of infection in sheep , goats , eattle and camels according to their sex .

	· · · · · · · · · · · · · · · · · · ·	Intensi	ty		
Examined animals	Sex	1 – 3 cysts (%)	4 – 8 cysts (%)	9 – 20 cysts (%)	p- value
Sheep	Males 73	39 (53,4)	27 (37.0)	7 (9.6)	- "
186	Females 113	61 (54.0)	40 (35.4)	12(10.6)	0.963
	Total 186	100(53.8)	67 (36.0)	19(10.2)	
	Males 105	66 (62.9)	31 (29.5)	8 (7.6)	
Goats 195	Females 90	54 (60.0)	25 (27.8)	11(12.2)	0.439
	Total 195	120(61.5)	56 (28.7)	19 (9.7)	
Cattle	Males 9	4 (44.5)	3 (33.3)	2 (22.2)	
17	Females 8	5 (62.5)	. 3 (37.5)	0 (0.0)	0.257
i	Total 17	9 (52.9)	6 (35.3)	2 (11.8)	
Camels	Males 7	2 (28.6)	3 (42.8)	2 (28.6)	
20	Females 13	6 (46.2)	4 (30.7)	3 (23.1)	0.559
	Total 20	8 (40.0)	7 (35.0)	5 (25.0)	

4.4 Hydatid cyst characteristics:

4.4.1 Cyst morphology:

Hydatid cysts in all animals examined were either superficial (plate 17a and b) or deeply (plate 18a and b) within the infected organs. Hydatid cysts in the liver were relatively easier to recognize visually than those in the lungs. The cyst walls were thick or thin and semi-transparent in nature.

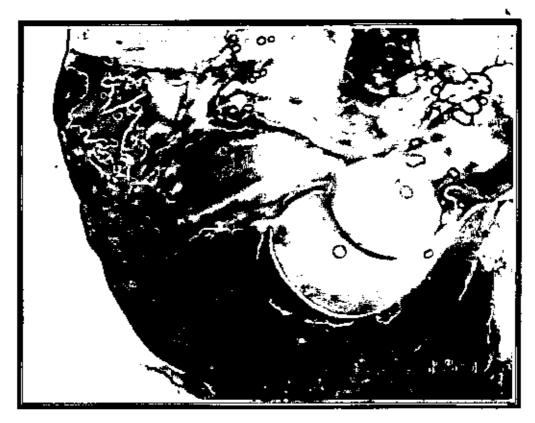
The shape of cysts seen was spherical, elliptical and / or irregular, whereas cysts of lungs were mostly spherical in shape. Cysts in sheep, cattle and camels were large cysts $(5-10\ {\rm cm})$ with inter-communicating chambers. The results showed that different shapes of hydatid cysts were seen, these cysts may be broadly grouped into two categories, ellipsoids and irregular shapes (plates 19 and 20).

4.4.2 Cyst dimensions:

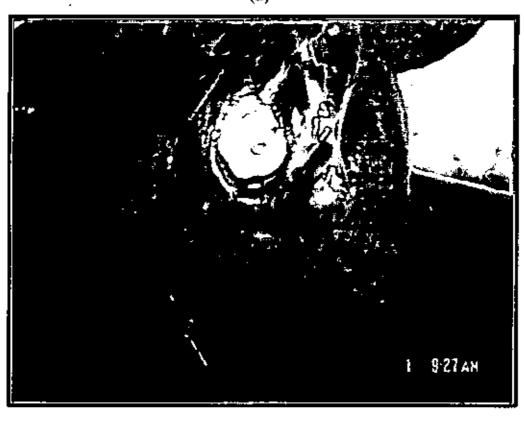
cyst dimensions for the four animal species examined are shown in Table 12. The size of hydatid cysts examined were various. The average of dimensions were between hairpin and 20 cm. The cysts were classified according to their size to four groups: small (less than 2 cm), medium (2-5 cm), large (5 - 10 cm) and very large (10 - 20 cm). In sheep most of hydatid cysts found in liver (56.7 %) were medium size (2 - 5 cm), whereas, those found in lungs (45 %) were small in size (<2 cm) (Table 13). In goats, 45.8 % of hydatid cysts in liver were medium size and 42.8 % in lungs were small size. Large hydatid cysts were found only in liver and lunges of goats as 4.2 % and 15.4 % respectively. Among all four species, the largest cysts were found in liver and lungs of goats. In cattle and camels, the most hydatid cysts were found in liver and lungs were small size (Table 14).

53

Results



(a)



(b)

Plate 17a, b: Hydatid cysts of Echinococcus granulosus in camel liver.

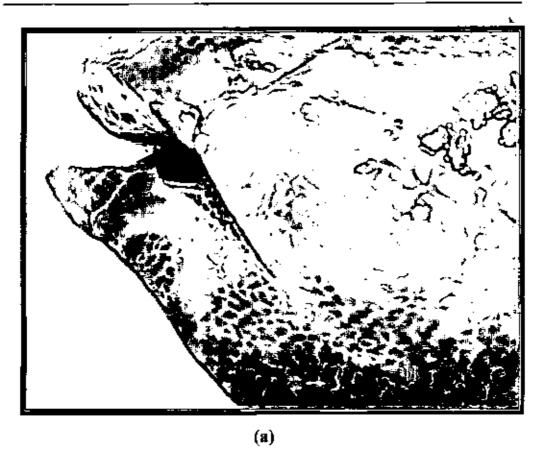


Plate 18 a , b : Hydatid cysts of Echinococcus granulosus in cattle liver .

(b)



Plate 19: Sheep liver infected with hydatid cysts.



Plate 20: Goat lungs infected with hydatid cysts.

Table 12: The size of hydatid cysts in liver and lungs of sheep, goats, cattle and camels.

!		No. of		Cyst	size	
Examined animals		cysts measured	<2cm (%)	2-5cm (%)	6-10cm (%)	11- 20cm (%)
Sheep	Liver	30	10(33.3)	17(56.7)	3 (10.0)	0 (0.0)
: !	Lungs	20	9 (45.0)	8 (40.0)	3 (15.0)	0 (0.0)
Goats	Liver	24	9 (37.5)	11(45.8)	3 (12.3)	1 (4.2)
	Lungs	26	11(42.3)	10(38.5)	1 (3.8)	4 (15.4)
Cattle	Liver	30	20(66.7)	4 (13.3)	6 (20.0)	0 (0.0)
	Lungs	20	10(50.0)	5 (25.0)	5 (25.0)	0 (0.0)
Camels	Liver	29	18(62.1)	6 (20.7)	5 (17.2)	0 (0.0)
ļ	Lungs	21	13(61.9)	5 (23.8)	3 (14.3)	0 (0.0)

Table 13: The size of hydatid cysts of sheep, goats, cattle and camels according to their sex.

	·			Cyst s	ize		
Examined animals	Sex	No. of cysts measured	<2cm (%)	2-5cm (%)	6-10cm (%)	11- 20cm (%)	p-value
Sheep	Males 25	25	6(24.0)	16(64.0)	3(12.0)	0 (0.0)	0.138
	Females 25	25	13(52.0)	9(36.0)	3(12.0)	0 (0.0)	
Goats	Males 22	22	6(27.3)	9(40.9)	4(18.2)	3 (13.6)	0.043*
	Females 28	28	14(50.0)	12(42.9)	0 (0.0)	2 (7.1)	0,043
Cattle	Males 32	32	17(53.1)	7(21.9)	8(25.0)	0 (0.0)	0.266
,	Females 18	18	13(72.2)	2(11.1)	3(16.7)	0 (0.0)	0.200 .
Camels	Males 21	21	12(57.1)	8(38.1)	1(4.8)	0 (0.0)	0.619
	Females 29	29	19(65.5)	3(10.3)	7(24.2)	0 (0.0)	5.017

Table 14: The size of hydatid cysts of sheep, goats, cattle and camels according to their age.

				Cyst	size		
Examined animals	Age (Years)	No. of cysts measured	<2cm (%)	2-5cm (%)	6-10cm (%)	11-20cm (%)	p-value
	<1	10	5(50.0)	3(30.0)	2(20.0)	0 (0.0)	
	1-<2	10	3(30.0)	5(50.0)	2(20.0)	0 (0.0)	
Sheep	2-<3	13	7(53.8)	5(38.5)	1 (7.7)	0 (0.0)	
	≥3	17	4(23.5)	12(70.6)	1 (5.9)	0 (0.0)	0.908
!	Total	50	19(38.0)	25(50.0)	6(12.0)	0 (0,0)	
	<1	12	6(50.0)	6(50.0)	0 (0.0)	0 (0.0)	
ļ	1-<2	9	3(33.3)	6(66.7)	0 (0.0)	0 (0,0)	
Goats	2-<3	14	5(35.7)	4(28.6)	4(21.4)	2(14.3)	
	≥3	15	6(40.0)	5(33.3)	1 (6.7)	3(20.0)	0.068
	Total	50	20(40.0)	21(42.0)	5(10.0)	5(10.0)	
	<j< th=""><th>10</th><th>9(90.0)</th><th>1(10.0)</th><th>0 (0.0)</th><th>0 (0.0)</th><th></th></j<>	10	9(90.0)	1(10.0)	0 (0.0)	0 (0.0)	
Cattle	1-<2	12	7(58.3)	5(41.7)	0 (0.0)	0 (0.0)	į
	2-<3	19	9(47.4)	1 (5.3)	9(47.4)	0 (0.0)	
	≥3	9	5(55.6)	2(22.2)	2(22.2	0 (0.0)	l l
	Total	50	30(60.0)	9(18.0)	11(22.0)	0 (0.0)	0.023*
	<1	6	6(100.0)	0 (0.0)	0 (0.0)	0 (0.0)	
Camels	1-<2	12	7(58.3)	2(16.7)	3(25.0)	0 (0.0)	
	2-<3	16	8(50.0)	7(43.8)	1 (6.2)	0 (0.0)	
	≥3	16,	10(62.5)	2(12.5)	4(25.0)	0 (0.0)	0.248
	Total	50	31(62.0)	11(22.0)	8 (16.0)	0 (0.0)	

4.4.3 Fertility of hydatid cysts:

The physical content of hydatid cysts may be fluid, caseous, or calcified. Sheep had significantly higher percentage of fertile cysts (72%) than other animals. Cattle had the higher percentage of sterile cysts among the four inspected species (70%). Calcified cysts accounted for 14% in cattle and in sheep and camels 4% each, while goats had 8%. On the other hand, caseous cysts were more frequently observed in cattle (16%) than that in other three species (8% for each) (Table 15 and Fig. 15).

The age groups had no effect on the cyst fertility in any animal (P < 0.05). There is no evidence indicate that age groups had species, effect on the fertility of hydatid cysts (Table 16). Among all animal species, the results shown that sex had no effect on cyst fertility (P > 0.05) (Table 17). The relationship between fertility and organ type is shown in Table (18). Liver of sheep and camels had more fertile cysts than the lungs but there was no statistical difference between cysts of the two organs (P > 0.05).

4.4.4 Viability of hydatid cyst protoscoleces:

The viability of hydatid cysts was determined through the relationship between the ratio of viable protoscoleces to total number of the protoscoleces in each cyst. The viability of hydatid cyst protoscoleces was classified according to their percentage into three categories: low viability (5 – 40 %), medium viability (41 – 60 %) and high viability (61 – 100 %). The results showed that high viability (38 %), medium viability (20 %) and low viability (12 %) of protoscoleces in sheep, whereas in goats 26 % high, 16 % for both medium and low viability. In camels, 32 % of viability was medium and 20 % and 18 % were low and high viability (Table 19).

Table 15: Classification of hydatid cysts of examined sheep, goats, cattle and camels according to their fertility and physical contents.

Hydatid cyst	Sheep	Goats	Cattle	Camels
categories	N (%)	N (%)	N (%)	N (%)
Fertile	36 (72.0)	29 (58.0)	0 (0.0)	35 (70.0)
Sterile	8 (16.0)	13 (26.0)	35 (70.0)	9 (18.0)
Calcified	2 (4.0)	4 (8.0)	7 (14.0)	2 (4.0)
Caseous	4 (8.0)	4 (8.0)	8 (16.0)	4 (8.0)
Total	50	50	50	50

X2 = 71.054 , P < 0.05 , df = 9 , p-value = 0.000 ***

Figure 15: Fertility of hydatid cyst from examined sheep, goats, cattle and camels.

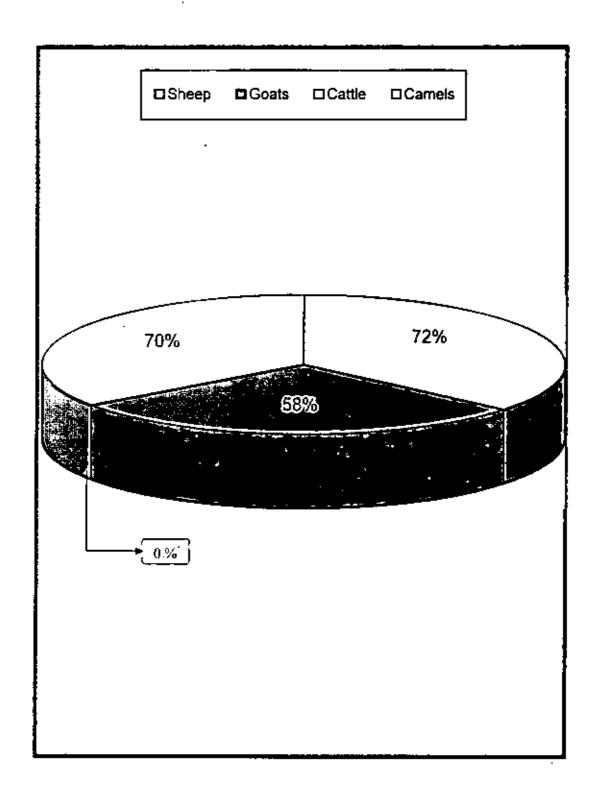


Table 16: The fertility of hydatid cyst of examined sheep, goats, cattle and camels according to their age.

Examined animals	Fertil	ity accordi	ng to age (ye	ears)	P-value
	<1 (%)	1-<2 (%)	2-<3 (%)	≥3 (%)	
Sheep	7 (70.0)	7 (70.0)	7 (53.8)	15 (88.2)	0.517
Goats	40 (33.3)	7 (77.8)	7 (50.0)	11 (73.3)	0.354
Camels	4 (66.7)	7 (58.3)	11 (68.8)	7 (70.0)	0.542

Table 17: The fertility of hydatid cysts of examined sheep, goats and camels according to their sex.

Hydatid cyst	SI	іеер	G	oats	Camels	
categories	Males	Females	Males	Females	Males	Females
No. of cysts	25	25	22	28	21	29
Fertility (%)	18 (72.0)	(72.0)	13 (59.1)	16 (57.1)	16 (76.2)	19 (65.5)
p-value	0.759 (Non-sig)	0.658 (Non-sig)	0.436 (Non-sig)	

Table 18: Comparsion between the fertility of hydatid cysts in liver and lungs of sheep, goats and camels.

		Fertile cysts								
Examined animals		Liver		Lungs						
	No.	Fortile	No.	Fertile	p-value					
Sheep	30	25 (83.3)	20	11 (55.0)	0.759					
Goats	24	12 (50.0)	26	17 (65.4)	0.521					
Camels	30	22 (73.3)	20	13 (65.0)	0.641					

Table 19: Comparsion between the viability of hydatid cyst protoscoleces from liver, lungs of sheep, goats and camels.

		Viabi	lity		I I h l .	
Examined animalsx	No. of cysts	5-40 %	41-60 %	61-100 %	Unviable	
Sheep	50	6 (12.0)	10 (20.0)	19 (38.0)	15 (30.0)	
Goats	50	8 (16.0)	8 (16.0)	13 (26.0)	21 (42.0)	
Camels	50	10 (20.0)	16 (32.0)	9 (18.0)	15 (30.0)	

4.4.5 Viability of cysts and age:

The results showed that there was no significant difference between viability of protoscoleces and age groups of all animal species, sheep (P = 0.244), goats (P = 0.833) and camels (P = 0.248) (Table 20).

4.4.6 Viability of cysts and sex:

There was no significant relationship between the viability of hydatid cyst and sex of infected sheep (P = 0.881) goats (P = 0.563) and camels (P = 0.187) (Table 21).

4.4.7 Viability of cysts and infected organs:

Fifty hydatid cysts of infected liver and lungs from sheep, goats and camels were examined for viability of protoscoleces. Both organs showed viable cysts, liver cysts showed 80 % and lungs 55 % in sheep, whereas 50 % and 65 % in liver and lungs of goats and liver and lungs of camels showed 76 % and 62 % of viable cysts respectively. The difference in the viability between two organs was insignificant (P>0.01) (Table 22).

4.4.8 Viability of cysts and cyst size:

Viable protoscoleces were detected in all sizes of hydatid cysts from sheep, goats and camels. There is a significant difference between the viability of protoscoleces and cyst sizes in sheep (P = 0.046) in goats (P = 0.042) and in camels (P = 0.048) (Table 23).

4.5 Measurement of protoscoleces:

Two hundred protoscoleces were examined and measured from each. organ (liver and lungs) of sheep, goats and camels (Table 24). Total length of protoscoleces of liver and lungs in sheep were found to be

Table 20: The viability of cyst protoscoleces of examined sheep, goats and camels according to their age.

			\	/iability %	6 (N)		
Examined animals	Age (Years)	No. of cysts	5-40 %	41-60 "%	61-100 %	Total viable cysis (%)	p- value
	<1	10	6(60.0)	1 (10.0)	0 (0.0)	7(70.0)	
Sheep	1-<2	10	0 (0.0)	4(40.0)	2(20.0)	6(60.0)	0.244
	2-<3	13	0 (0.0)	1 (7.7)	6(64.2)	7(53.8)	
	≥3	17	0 (0.0)	4 (23.5)	11(64.7)	15(88.2)	
	<1	12	2(16.7)	2(16.7)	0 (0.0)	4(33.3)	
Goats	1-<2	9	5(55.6)	2(22.2)	0 (0.0)	7(77.8)	0.833
:	2-<3	14	0 (0.0)	4(28.6)	3(21.4)	7(50.0)	
	≥3	15	1 (6.7)	0 (0.0)	10(66.7)	11(73.3)	
	<1	10	6(60.0)	1 (10.0)	0 (0.0)	7(70.0)	
Camels	1-<2	10	0 (0.0)	4(40.0)	2(20.0)	6(60.0)	0.248
	2-<3	13	0 (0.0)	1 (7.7)	6(46.2)	7(53.8)	
	≥3	17	0 (0.0)	4(23.5)	11(64.7)	15(88.2)	

Table 21: The viability of cyst protoscoleces of examined sheep, goats and camels according to their sex.

			 .	Viabilit	У		,
Examined animals	Sex	No. of cysts	5-40 %	41-60 %	61-100 %	Total viable cysts %	p- value
Sheep	Males	25	1 (4.0)	7(28.0)	10(40.0)	18(72.0)	
	Females	25	5(20.0)	3(12.0)	9(36.0)	17(68.0)	0.881
Goats	Males	22	4(18.2)	5(22.7)	4(18.2)	13(59.0)	
	Females	28	4(14.3)	3(10.7)	9(32.1)	16(57.0)	0.563
Camels	Males	21	6(28.6)	7(33.3)	3(14.3)	16(76.0)	
	Females	29	4(13.8)	9(31.0)	6(20.7)	19(65.5)	0.187

Table 22: The viability of hydatid cyst protoscoleces in liver and lungs of examined sheep, goats and camels.

			Via	bility %	(N)	•	
Examined animals	Location	No. of cysts	5-40 %	41-60 %	61-100 %	Total viable cysts %	p- value
Sheep	Liver	30	4(13.3)	5(16.7)	15(50.0)	24(80.0)	0.350
 	Lungs	20	2(10.0)	5(25.0)	4(20.0)	11(55.0)	
Goats	Liver	24	3(12.5)	2(8.3)	7(29.2)	12(50.0)	0.170
	Lungs	26	5(19.2)	6(23.1)	6(23.1)	17(65.0)	
Camels	Liver	29	6(20.7)	9(31.0)	7(24.1)	22(76.0)	0.658
	Lungs	21	4(19.0)	7(33.3)	2(9.5)	13(62.0)	

Table 23: The viability of cyst protoscoleces in examined sheep, goats and camels according to the cyst size.

	Cyst	No.		Viab	ility		
Examined animals	size	of cysts	5-40 %	41-60 %	61-100 %	Total ' viable cysts %	p- value
	<2 cm	19	3(15.8)	3(15.8)	0 (0.0)	6(31.6)	0.046*
Sheep	2-5 cm	25	3(12.0)	6(24.0)	15(60.0)	24(96.0)	
:	6-10cm	6	0 (0.0)	1(16.7)	4(66.7)	5(83.3)	
	11-20cm	20	3(15.0)	1 (5.0)	2(10.0)	6(30.0)	
 	<2cm	21	5(23.8)	5(23.8)	6(28.6)	16(76.2)	
Goats	2-5cm	4	0 (0.0)	1(25.0)	2(50.0)	3(75.0)	0.042*
	6-10cm	5	0 (0.0)	1 (20.0)	3(60.0)	4 (80.0)	i
	11-20cm	19	3(15.8)	3(15.8)	0 (0.0)	6 (31.6)	
	<2cm	31	10(32.3)	11(35.5)	0 (0.0)	21(67.7)	
Camels	2-5cm	11	0 (0.0)	3(27.3)	4(36.4)	7(63.6)	0.048*
!	6-10cm	8	0 (0.0)	2(25.0)	5(62.5)	7(87.5)	
	11-20cm	31	10(32.3)	11(35.5)	0 (0.0)	21(67.7)	

132.7-226.9 μm (Ave. 180.5) and 103.8 – 186.8 μm (Ave. 171.3 μm) respectively and in goats the measurement of protoscoleces were 120.6 – 256.7 μm (Ave. 181.1μm) and 113.5 – 272.7 μm (Ave. 174.8 μm) in liver and lungs , whereas in camels were 171.2 – 334.7 μm (Ave. 235.2 μm and 230.3 – 253.1 μm (Ave. 237.2 μm) . The width of protoscoleces from liver and lungs were 92.3 –238.6 μm (Ave.145.2μm) and 95.7 – 226.8 μm (Ave. 145.7μm) in sheep , 97.2 – 216.4 μm (Ave. 155.9 μm) and 102.2 – 215.5 μm (Ave. 150.9 μm) in goats whereas 124.6 – 277.4 μm (Ave. 195.1μm) and 103.2 – 282.9 μm (Ave. 202.2 μm) in camels respectively .

Characteristic arrangement of rostellum hooks was clearly seen in viable cysts from liver and lungs and the total number of hooks perprotoscoleces were found 28-38 (mean 32.6) in sheep ,28-42 (mean 33.9) in goats and 26-41 (mean 34.1) in camels. There is no differences between the number of hooks of liver and lung cyst in each animal species and between sheep , goats and camels .

4.5.1 Rostellum hooks:

The dimensions of small and large hooks are shown in Table (25). Differences between hooks in both liver and lungs were detected. Among large hooks the average total length was 23.9 μm , 22.8 μm and 24.5 μm , blade length was , 13.6 μm , 14.1 μm and 11.7 μm , handle length was 9.9 μm , 9.2 μm and 9.9 μm , hook's width was 7.4 μm , 8.1 μm and 8.3 μm in sheep , goats and camels respectively . While small hooks showed average total length 19.6 μm , 20.9 μm and 20.9 μm , blade length 11.7 μm , 12.7 μm and 12.3 μm , handle length 8.3 μm , 8.3 and 7.3 μm , hook's width was 7.2 μm , 7.2 μm and 6.4 μm in sheep , goats and camels respectively . There were no significant differences between rostellum hooks measurements and animal species (Plate 21) .

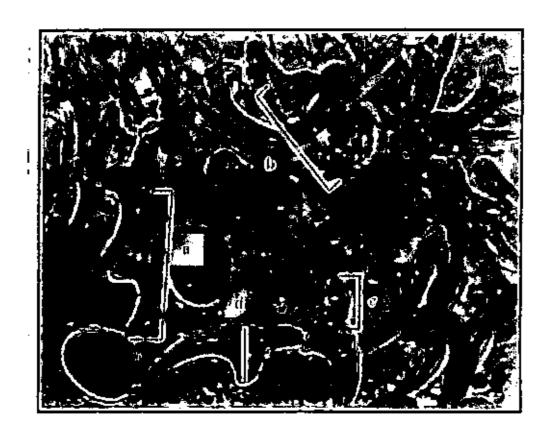


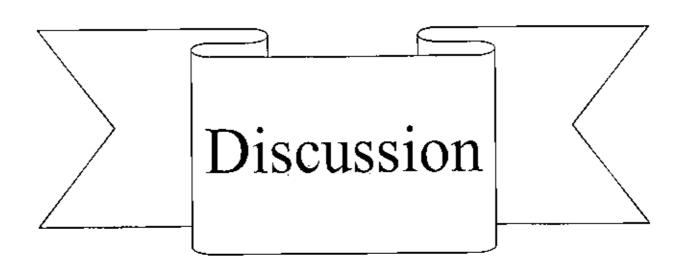
Plate 21: Rostellum hooks from cyst showing measurement recorded $X=100\ (a)\ Total\ length\ , (b)\ Blade\ length\ , (c)\ Handle\ length\ , (d)\ Hook\ width\ .$

Table 24: Measurement of protoscoleces and total number of their hooks from liver and lungs of examined sheep, goats and camels.

		No. of cysts	No. of	Mea	surement	Measurement of protoscoleces	ş	New See as I work	l sook
Examined animals	Organ	examined	protoscoleces examined	Total length (µm)	յքի (բա)	Total width (µm)	(աղ) կ	scolex	×
				Range	Mean	Range	Mean	Range	Mean
Sheep	Liver	25	200	132.7-226.9	180.5	92.3-238.6	145.2	28-38	32.9
	Lungs	25	200	103,8-186.8	171.3	95.7-226.8	145.7	28 – 38	32.2
Goats	Liver	2.5	200	120.6-256.7	181.1	97.2-216.4	155.9	28 – 42	33.1
· · · · · · · · · · · · · · · · · · ·	Sanrı	25	200	113.5-272.7	174.8	102.2-215.5	150.9	29-41	34.6
Camels	Liver	25	300	171.2-334.7	235.2	124.6-277.4	195.1	26-41	34.1
	[rangs]	25	200	230.3-253.1	237.2	103.2-282.9	202.2	25 – 40	34.2

Table 25: Measurements of rostellum hooks from hydatid cysts of sheep, goats and eamels.

Animal type Rostellar hooks	Sheep	Goats	Camels
Large hooks	N = 200	N = 200	N = 200
Hook's total length	23.9 (21.2 – 25.5) µm	22.8 (20.1 – 27.3) µm	24.5 (23.1 – 26.6) µm
Blade length	13.6 (14.3 – 16.1) µm	14.1 (10.8 – 16.7) µm	11.7 (10.7 – 15.9) µm
Handle length	9.9 (7.8 – 11.1) µm	9.2 (8.4 – 10.8) µm	9.9 (8.2 – 11.6) µm
Hook's width	7.4 (6.5 – 8.6) µm	8.1 (7.3 – 9.7) tun	8.3 (6.3 – 10.2) µm
Small hooks	N = 200	N = 200	N = 200
Hook's total length	19.6 (18.5 – 23.9) µm	20.9 (18.6 – 25.2) µm	20.9 (18.1 – 22.8) µm
Blade length	11.7 (10.4 – 15.1) µm	12.7 (10.8 – 15.6) µm	12.3 (9.5 – 15.3) µm
Handle lenoth	8.3 (7 – 10.6) µт	8.3 (7.3 – 9.8) µm	7.3 (5.5 – 8.4) µm
Hook's width	7.2 (6.5 – 9.9) µm	7.2 (7.5 – 8.7) μm	6.4 (5.2 – 9.1) µm



5. DISCUSSION

This study was carried out to display the prevalence of echinococcosis among domestic animals at Sirt city. From the epidemiological point of view echinococcosis is a cyclozoonotic infection having a worldwide distribution and a variable geographic incidence (Williams et al., 1971). The incidence in any country is closely related to prevalence of the disease in domestic animals (Craig et al., 1995 and Gottstein, 2000).

It is well established that hydatid disease is an important health problem and livestock economy in Libya (Dar and Taguri, 1979; Gebreel et al., 1983; Gusbi et al., 1987 and Shaafie et al., 1999). The prevalence of hydatid cysts in intermediate hosts provides an indicator of all degree of pasture contamination with *Echinococcus* eggs released in dog faeces (Schwabe, 1968).

5.1 Prevalence:

The hydatid cysts of *E. granulosus* were detected in all examined animal. These animals included sheep, goats, cattle and camels. This finding are in agreement with previous studies in Libya (Gebreel *et al.*, 1983; Gusbi *et al.*, 1987; Ahmed *et al.*, 1996; Ibrahem and Gusbi, 1997; Al-Saqur *et al.*, 2001), Aboudaya (1985a); Tashani *et al.* (2002) and Mohamed *et al.* (2004).

5.1.1 Sheep:

The results have showed that the prevalence of hydatid cysts in slaughtered sheep was found to be 4.9 % in Sirt. This result came in agreement with the prevalence rates reported from different localities in Libya, the incidence of hydatidosis in sheep during 1975 and 1977 was

3.1% and 3.3 % respectively (Dar and Taguri, 1979). Other study for the same period revealed that 3.08 % and 3.34 % of slaughtered sheep were infected (Gebreel et al., 1983). Aboudaya (1985a) collected data of hydatid cysts slaughtered sheep in 46 abattoirs distributed throughout Libya and he found the prevalence in sheep was 4.3 %. The lower prevalence of hydatidosis in examined sheep seen in the present study have been reported from other countries, southern Morocco (5.3%) (Pandey et al., 1988), central Jordan (1.3 %) (Dajani and Khalaf, 1981), central Iraq (5.9 %) (Al-Abbassy et al., 1980), northwestern Morocco (0.7 %) (Pandey et al., 1986).

While the high prevalence in sheep throughout the country was done by Gusbi et al. (1987) given an overall prevalence at 12.74 %. Ibrahem and Craig (1998) reported that 15.6 % of slaughtered sheep from Benghazi were infected with hydatidosis. In a recent study, Al-Saqur et al. (2001) reported that 14.3 % of sheep were infected with hydatid cysts in Sebha. The prevalence of hydatid disease in sheep from Benghazi was found to be 20 % (Tashani et al., 2002) and from Sebha was 18.5 % (Mohamed et al., 2004) and from other countries, from north and central Tunisia (65.6 %) (Lahmar et al., 1999), southern Morocco (44.6 %) (Pandey et al., 1988) northern Jordan (28 %) (Abdel-Hafez et al., 1986a), northwestern Iran (19.1 %) (Nourian et al., 1997) and northern Iraq (15 %) (Saced et al., 2000).

5.1.2 Goats:

The prevalence of hydatidosis in examined goats from Sirt was found to be 2.4 %. This result is consistent with those reported from other regions of Libya, from Benghazi was reported to be 5.5 % (Dar and Taguri, 1979), 5.5% (Ibrahem and Craig, 1998), 3.4% (Tashni et al.,

2002). Gusbi et al. (1990) reported that the infection was 1.5 % throughout the country. The infection were 8.2 % and 5.8 % in goats from Sirt and El-Kumes respectively (Ibrahem and Craig, 1998) and 4% from Sebha (Mohamed et al., 2004). Similarly low infection rates of hydatidosis in goats have been reported from other countries, Syria (2.3%) (Dajani, 1978), central Iraq (5.1 %) (Al-Abbassy et al., 1980), central Jordan (0.54 %) (Dajani and Khalaf, 1981), northern Jordan (3.6%) (Al-Yaman et al., 1985), Kenya (7.1 %) (Macpherson, 1985), northwestern Morocco (1.4 %) (Pandey et al., 1986), India (1.9 %) (Irshadullah et al., 1989) and northern Iraq (6.2 %) (Saced et al., 2000).

High infection was reported from Benghazi at 18.1 % (Gebreel et al., 1983). The low infection rate in goats may due to the feeding behaviour of this animal, they feed on the upper parts of plants which decrease the chance of these animals to ingest the *Echinococcus* eggs, which usually on the pasture and lower parts of plants, this was reported by Cousi (1951), Pandey et al. (1986), Euzéby (1991), Tashani et al. (2002) and Mohamed et al. (2004).

5.1.3 Cattle:

Prevalence in cattle in the present study was 15 %, this result was significantly higher than that in sheep, goats and camels. The prevalence of hydatidosis in cattle in Benghazi during the years of 1975, 1976 and 1977 was 8.9 %, 10.6 % and 13.9 % respectively Dar and Taguri (1979) and Gebreel et al., 1983). while a nationwide study revealed that the prevalence of hydatidosis in cattle was found to be from 0.7 % to 37 % (Aboudaya, 1985a). Gusbi et al. (1990) recorded that infection rate in cattle from Tripoli was 5.7 %. The prevalence rate of E. granulosus in cattle from Benghazi was 11 % (Tashani et al., 2002). On the other hand

Mohamed et al. (2004) reported the same prevalence obtained in the present study in cattle (15 %) from Sebha!

Similar and different infection rates in cattle have been recorded from various parts of the world. The previous studies revealed that high infection rates were recorded from Bangladesh (42.15 %) (Islam, 1982), Morocco (44.6 %) (Pandey et al., 1988) and northwestern Iran (22 %) (Nourian et al., 1997). On the other hand low prevalence rates of hydatidosis in cattle were reported from Kano state, Nigeria (14.7 %) (Dada et al., 1980), central Iraq (4.9 %) (Al-Abbassy et al., 1980), northern Jordan (11.4 %) (Al-Yaman et al., 1985), Kenya (8.9 %) (Macpherson, 1985), northern Iraq (10.9 %) (Saeed et al., 2000).

5.1.4 Camels :

The prevalence of hydatidosis in examined camels was 2.7 %. This result came in agreement with those the results obtained by Dar and and Taguri (1979) at 1.41 %, Gebreel et al. (1983) at 1.48 %, Khan and El-Buni (1999) in Benghazi at 1.97 % and Gdourra (2003) in Sirt at 3.62 %. Dar and Taguri (1979) reported that the prevalence rate in camels during the years 1976 and 1977 was 34.7 % and 29.10 % respectively. During the same years Gebreel et al. (1983) made a study on Echinococcosis in Libya and revealed that the prevalence was detected at 17.6 % and 29.1% respectively. However, many reports shows that hydatidosis in camels was highly endemic in Libya 27.20 % (Aboudaya, 1985a), 5.9 % (Gusbi et al., 1987, 1990), 48.4 % in Sebha (Ahmed et al., 1996), 48 % (Ibrahem and Craig, 1998), 13.6 % (Tashani et al., 2002) and 12.62 % (Mohamed et al., 2004). On the other hand high prevalence of hydatidosis in camels was reported in Kufra at 50.6% (Gusbi et al., 1990).

Ibrahem and Craig (1998) reported that the prevalence of hydatidosis in camels from six slaughter houses in northern Libya was found to be 48 %, the prevalence in these six localities were 55 % in Mesurate, 49.5 % in Sirt, 47.5 % in Tripoli, 42.4 % in El-Khumes, and 38.7 % in Zawia. Gusbi et al. (1990) reported that the prevalence was 36.2 % in camels from Benghazi . The high infection prevalence rates of hydatidosis in camels included in the previous studies may due to those camels were brought from various regions within the country in addition to those imported camels to Libya from its neighbours such as Sudan, Tunisia, Niger and Chad, where hydatidosis had been reported in these countries, 45.4 % in camels from Sudan (Saad et al., 1983), 66 % in Tunisian camels (Ben Osman, 1965) and 22 % in Niger camels (Develoux et al., 1985). In Egypt, Haridy et al. (1998) among camels imported from Sudan for human consumption over five years reported 5.5 % (1992) , 6.1 % (1993) , 6.7 % (1994) , 8.2 % (1995) and 4.3 % (1996) . On the other hand low infection rate recorded may explained by those camels are brought to slaughter from restricted area specially around Sirt or due to the large number of young camels in the examined samples .

Hydatid disease in camels have been reported from other countries, these results may be similar or higher than that seen in Libya, such results were 20.4 % in central Iraq (Al-Abbassy et al., 1980), 8.8 % in northern Jordan (Al-Yaman et al., 1985) and high infection a 80 % in Morocco (Pandey et al., 1986). Generally speaking, in various African countries, camels (C. dromedaries) was infected with hydatid cysts of E. granulosus. This recommended that camels are reservoir for humans infection (Hamdy et al., 1980 and Macpherson et al., 1986).

5.1.5 Prevalence and sex:

Both sexes of all animal species examined in the present study were found to be infected with hydatidosis. No significant differences were found between infected male and female sheep. Both sexes have shown nearly the same rate of susceptibility to the infection. This disagree with other authors, who found that females sheep were infected more than males (Pandey et al., 1988 and Tashani et al., 2002).

The prevalence rate in both male and female examined sheep in the present study may explain the susceptibility of both sexes to infection. On the other hand hydatidosis was more prevalent in females than males goats. While no significant differences were shown between both sexes of cattle. The result revealed that there was a significant difference of hydatidosis between sexes of camels. Al-Yaman et al. (1985) reported that female camels from northern Jordan were significantly more infected than males. A study conducted by Mohamed et al. (2004) in Sebha revealed that no significant differences were found between infected male and female and both sexes have shown the same susceptibility to infection with E. granulosus.

5.1.6 Prevalence and age:

The prevalence rates were increased with age in all examined animal species. This finding was demonstrated in many animal species (Al-Yaman et al., 1985; Pandey et al., 1986; Roberts et al., 1986 and Tashani et al., 2002).

The older sheep had higher prevalence than young. The same situation was seen in sheep for the prevalence of adult sheep in Libya (Gusbi et al., 1987 and Tashani et al., 2002), in Tunisia (Lahmar et al.,

correlation and significant difference between age groups of goats. An increase of prevalence with age was shown to occur in camels (Ibrahem and Craig, 1998; Tashani et al., 2002 and Mohamed et al., 2004). This finding disagree with results obtained by Gdourra (2003), the results showed that the infection of camels with hydatid cysts did not increase with age. The increase of infection rates with age may due to the older animals passed through a longer period of exposure to Echinococcus eggs and slow development of hydatid cysts take a longer time and easily detected in old animals. This situation have be reported in previous studies (Al-Abbassy et al., 1980; Islam, 1982; Al-Yaman et al., 1985; Pandey et al., 1988; Irshadulla etal., 1989 and Abo-Shehada, 1993).

5.1.7 Prevalence and seasons:

The season have no effect on the infection rate of hydatid cysts in all examined animal species. This may explained by those animals exposed to the *Echinococcus* eggs at the same degree over the year. Dyab et al. (2005) reported that prevalence of hydatid cysts in Egypt during Summer or Autumn was higher than during Winter or Spring.

5.1.8 Location of hydatid cysts:

The most commonly infected organs in all examined animal were liver and lungs. Liver was the most predominate site for hydatid cysts in sheep, goats and cattle. A similar findings reported in Libya (Gusbi et al., 1987, 1990; Tashani et al., 2002), in Middle East and Mediterranean, countries in north Jordan (Al-Yaman et al., 1985), in Tunisia (Lahmar et al., 1999), in northern Iraq (Saeed et al., 2000). On other hand, the lung was more infected than liver in sheep, cattle, goats and camels from Morocco (Pandey et al., 1988), Egypt (Haridy et al., 2000).

The lungs was found to be the most infected organ in the camels. This phenomenon was also reported from Jordan (Al-Yaman et al., 1985), Morocco (Pandey et al., 1986), Libya (Gusbi et al., 1990), Egypt (Abou-Aisha, 1999 and Haridy et al., 1998, 2000) This disagree with the results obtained by Gdourra (2003) in Sirt , the results showed that the liver was the most infected site for hydatid cysts in camels. It seems that the narrow size of lung capillaries and sponge texture of its tissue pave the way for the onchospheres to develop into cysts (Dyab et al., 2005) . Thompson et al., (1995) mention that an atomical structure and physiological characters of the host as well as the species and strain of parasite may explain the locations of cystic echinococcosis in various organs of intermediate hosts. On the other hand, Muller (2002) suggested that the highly lung infection with hydatid cysts my due to the possibility of infection occurs directly through the respiratory tract from Echinococcus eggs blown in dust by the wind. This situation may occurs in Libya.

The present study revealed that the sex had no effect on location of hydatid cysts in sheep, cattle and camels, while there is a significant difference between sex of goats and location of hydatid cysts. However age had effect on the location of cysts in sheep and goats, but such effect do not detected in cattle and camels. Harris et al. (1989) mentioned that the location of onchospheres may had further influenced by the age of animal at the time of exposure to infection.

5.1.9 Intensity of infection:

The majority of all examined animal showed light intensity of infection. This in agreement with many previous results (Al-Yaman et al., 1985; Pandey et al., 1986; Gusbi et al., 1990; Ibrahem and Craig,

1998; Tashani et al., 2002 and Mohamed et al., 2004). The intensity of infection in different organs of various intermediate hosts have been reported from different countries, from northern Jordan (Al-Yaman et al., 1985 and Abdel-Hafez et al., 1986b) and from Morocco (Pandey et al., 1986; 1988) in both countries the results revealed that the intensity was higher in camel lungs. On the other hand Ibrahem and Craig (1998) reported that the intensity of infection increase with animal age.

5.2 Hydatid cyst characteristics.

5.2.1 Size of hydatid cysts:

The size of hydatid cysts for examined animal species are varied in dimensions. The dimension of cysts in the liver and lungs of sheep, cattle and camels ranged from <2 cm to a maximum 10 cm, while in goats from <2 cm to a maximum 20 cm. Tashani et al. (2002) reported that the dimensions of cysts in the liver and lungs of sheep were similar and the same observations were seen in goats and camels cysts. In the present study infected goats had the largest hydatid cysts. This finding was observed in cattle and camels (Tashani et al., 2002). The size of hydatid cysts is controlled by the organ in which it grows (Gusbi et al., 1991).

The result showed that the cyst size increase with age in sheep, which agrees with the results obtained by Morris and Richards (1992); Ibrahem and Craig (1998) and Tashani et al. (2002). On the other hand, no relation between the size of the hydatid cysts and age in cattle was seen in Morocco (Pandey et al., 1988). The cyst size and age was poorly correlated in the case of cattle and camels, however goat cysts decrease in size with age increase (Tashani et al., 2002).

5.2.2 Fertility of hydatid cysts:

The percentage of fertility of sheep, goats, cattle and camels were 72 %, 58 %, 0.00 % and 70 % respectively. Sheep had the lowest sterile cysts (16 %) followed by camels (18 %) and goats (26%) while in cattle .(70%). This observation is consistent with results previously reported in 1.ibya, in sheep (69 %), cattle (14 %) and camels (51.7 %) respectively (Tashani et al., 2002). Haridy et al. (1998) reported that fertility was 49 % in camels from Egypt, and Wilson and Rausch (1980) put the fertility at 92 % in sheep and 80 % in cattle. High hydatid cyst fertility in camels was reported in northern Africa (Abdul-Salam and Farah, 1988; Gusbi et al., 1987; 1990; Ibrahem and Craig, 1998).

The examined livers of sheep and camels had more fertile cysts than lungs. In Egypt, Haridy et al. (1998) reported a fertility of 29 % in cysts of lungs and 20 % in liver cysts of camels. In sheep slaughtered in Egypt, Haridy et al. (2000) reported that the cysts in the lungs and the liver of sheep showed a fertility of 50 % and 40 % respectively. Ibrahem and Craig (1998) showed camel cysts (99.6 %) more fertile than sheep cysts (86.3 %) without mention the percentage in infected organs in the same study they reported that 71.4 % of fertile liver cysts and zero lung infection in goats. Many previous studies reported differences in fertility of liver and lungs hydatid cysts in intermediate hosts from different countries (Al-Abbassy et al., 1980; Pandey et al., 1988; Irshadulla et al., 1989; Mohamed et al., 1997; Abou-Aisha, 1999 and Khan et al., 2001).

5.2.3 Viability of hydatid cyst protoscoleces:

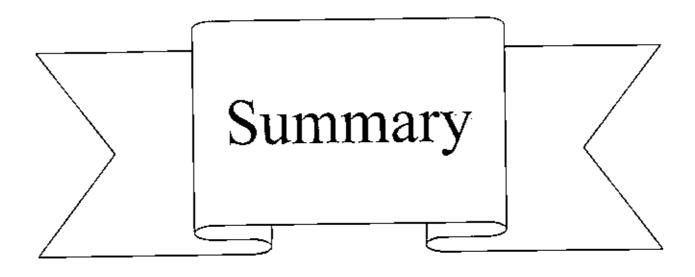
The viability of hydatid cysts in the sheep and camels was similar (70%). While in goats was lower (58%). The viability of sheep liver cysts (83.3%) was higher than lung cysts (55%), the same situation for camels, the viability of liver cysts was 73.3% and lungs cysts was 65%. However goat lung cysts (65%) was higher than that in the liver (50%). Irshadulla et al. (1989) and Mohamed et al. (2004) reported that the viability of camel lung cysts was higher than that in liver. Himonas et al. (1994) reported the same result in cattle in Greece. However, Tashani et al. (2002) reported that sheep liver cysts are more viable than lung cysts, while in camels, the viability of camel cysts (76%) was higher than that of lung cysts (45.5%).

The absence of reports on percentage of viability of *E. granulosus* cysts in intermediate hosts in Libya made the comparisons to be difficult. El-Sageyer and Kidwai (1993) reported the viability of hydatid cysts in sheep in Benghazi. The viability of hydatid cysts in sheep increased with age. The viability of cysts increased with increasing the cyst size in camels. While in sheep and goats the viability decreased with cyst size increased. Abdel-Hafez *et al.* (1986b) reported that the viability increased with increasing the cyst size in sheep. No significant influence of sex on the viability of cysts in sheep, goats and camels (P=0.881). This finding is in agreed with results reported by Abdel-Hafez *et al.* (1986b) and Mohamed *et al.* (2004).

5.2.4 Measurements of protoscoleces and rostellar hooks:

The size of protoscoleces from sheep and goats nearly similar, while those from camels quite different, and larger than protoscoleces from sheep and goats. There was no significant difference in the size of

rostellum hooks from liver and lungs in various examined animal species, the same finding was reported by Tashani *et al.* (2002). Variations in hook measurments was reported by Daily and Sweatman (1965). Many studies have been done to characterize the strain of *E. granulosus* by using hook morphology alone were not successful (Hobbs *et al.*, 1990). Constantine *et al.* (1993) and El-Sageyer and Kidwai (1993).



SUMMARY

A total of 3794 sheep, 8123 goats 113 cattle and 739 camels slaughtered at Sirt abattoir were examined for hydatid cysts of *Echinococcus granulosus*. The prevalence of hydatid cysts in these animals was found to be 4.9 %, 2.4 %, 15 %, and 2.7 % respectively.

The infection with hyadtid cysts was detected in both males and females of all examined animal species. The result showed that the prevalence in male sheep was 5.9 % and female was 4.4 %. No significant differences in prevalence of hydatidosis between both sexes (P= 0.58). The prevalence in females goats (3.9 %) was higher than in males (1.8 %) (P=0.000). The prevalence in male and female cattle was 12.9 % and 18.6 % respectively. No significant differences between both sexes of cattle (P = 0.411). However there was a slight correlation and significant difference (P = 0.044) in prevalence of hydatidosis between male (1.7 %) and female (4.1 %) camels.

The infection with hydatidosis was age dependent in sheep and goats, the infection increase with age. There are significant differences between the prevalence and age groups in sheep (P=0.000) and goats (P=0.000). Prevalence in cattle older than 3 years was 22.7 % higher than 1 year old (2.1 %) and 1-2 years old (14.3 %). The infection with hydatid cysts in camels increased with age. The differences between age groups were statistically significant (P = 0.000).

The results showed that there is no correlation and significant difference in prevalence and seasons (winter, spring, summer and Autumn).

Single infection was recorded in liver (51 %), lungs (22.6 %) and mixed was (26.3 %) in sheep. In goats the single infection was 39 % in liver. 27.7% in lungs and mixed infection was 33 % in both organs together. Infection in camel lungs (50%) was higher than liver (25 %), while infection in both organs was 25 %. In sheep liver infection (77.9 %) was higher than lungs (48.9 %). Goats liver had 72.3 % of infection and lungs 60 %. Cattle liver infection was 64.7 % and lungs 58 %. However, lungs of camels (75 %) more infected than liver (50 %).

The sex had no effect on location of cysts in sheep, cattle and camels. However, there is a significant difference between sex of goats and location of cysts (P=0.011). There was a significant difference between age and location of cysts in sheep and goats (P=0.000), but this finding do not detected in cattle and camels.

Intensity of infection was correlated and a significant in different age groups in sheep and goats (P = 0.000), but it is not in cattle (P = 0.699) and camels (P = 0.053). The sex of examined animals had no effect on the intensity of infection.

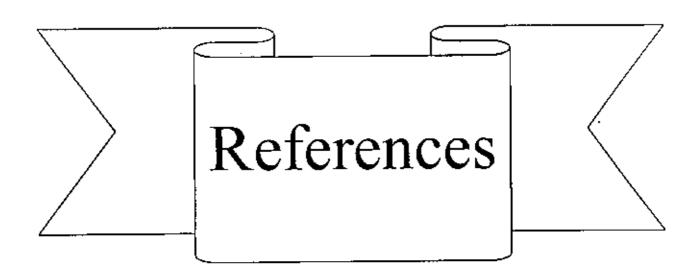
The most of hydatid cysts in sheep were found in liver (56.7 %) in the size (2-5 cm), whereas, those found in lungs (45.1 %) medium small size (> 2cm). 45.8 % of goats liver had medium size, while 42.8% of lungs had small size. Large hydatid cysts (5-10 cm) were found only in liver (4.2 %) and lungs (15.4 %) of goats. In cattle and camels, the most hydatid cysts of small size were found in liver and lungs.

The physical content of hydatid cysts was fluid, caseous and classified. The fertility of hydatid cysts in examined animals were 72 %, 58 %, 0.00 % and 70 % in sheep, goats, cattle and camels respectively. All cysts of cattle were sterile. The sheep and camels liver had more fertile cysts than lungs. Age groups had no effect on the cyst fertility in each examined animal species (P > 0.05).

There was no correlation between viability of hydatid cysts and age or sex of sheep , goats , cattle and camels . The differences in the viability between liver and lungs of, all examined animals were insignificant (P=0.01) . However there is a significant difference between viability and cyst size in sheep (P = 0.046) , goats (P = 0.042) and camels (P 0.048) .

The average length of protoscoleces of liver in sheep , goats and camels were 180.5 μm , 181.1 μm and 235.2 μm respectively . Large protoscoleces were noted among those isolated from hydatid cysts of camels .

Characteristic arrangement of rostellar hooks was clearly seen in viable cysts from liver and lungs. There was no differences between the number of hooks in liver and lung hydatid cysts in each animal species and between sheep, goats and camels. There were no significant differences between rostellar hooks measurements in the examined animal species.



REFERENCES

Abdel-Hafez, S. K.; Al-Yaman, F. M. and Said, I. M. (1986a). Further studies on prevalence of hydatidosis in slaughtered animals from North Jordan. Zeitschrift für Parasitenkunde, 72:89-96.

Abdel-Hafez, S. K.; Said, I. M. and Al-Yaman, F. M. (1986b). Comparative aspects on the fertility and viability of hydatid cysts in sheep from North Jordan. *Japanese Journal of Parasitology*, 35: 491-496.

Abdul-Salam, H. M. and Farah, M. A. (1988). Hydatidosis in camels in Kuwait. *Parasitology Research*, 74:267-270.

Abo-Shady, A. F. (1980). Intestinal helminthes among stray dogs in Mansoura city, Egypt. Journal of the Egyptian Society of Parasitology 10:289-294.

Abo-Shehada, M. N. (1993). Some observations on hydatidosis in Jordan. Journal of Helminthology, 67:248-252.

Aboudaya, M. A. (1985a). Prevalence of human hydatidosis in Tripoli region of Libya. *Garyounis Medical Journal*, 9:307-310.

Aboudaya, M. A. (1985b). Prevalence of Echinococcus granulosus among domestic animals in Libya. Tropical Animal Health and Production, 17:169-170.

Abou-Eisha, A. M. (1999). Prevalence of hydatid cysts in slaughtered animals in relation to public health. 5th Scientific Congress, Egyption Society for Cattle Diseases, 28 – 30 November.

Achour, N.; Dammak, J.; Zouari, B.; Nacef, T.; Belaid, A.; Mestiri, S. and Farhat, M. (1989). Epidémiologies du kyste hydatique en Tunisie. (A propos de 4124 dossiers de maladies operés entre 1977 et 1982). La Tunisie Medicale 66: 21 – 26.

Adams, F. (1849). The Genuine works of Hippocrates. *The Sydenhan Society*, London.

Ahmed, L. S. (1991). Incidence of hydatid disease in camels slaughtered at Assiut abattoir. *Assiut Veterinary Medical Journal*, 24: 274-278.

Ahmed, M. B.; Fathelbab, G. O.; Saffor, K. M.; Soum, S. A. and Ibrahem, S. A. (1996). Study for Hydatidosis in slaughtered camels in central abattoir of Sebha. *Journal of the Libyan veterinary sciences society*, Vol. 1, No. 1, 2:38-50.

Al-Abbassy, S. N.; Altaif, K. I.; Jawad, A. K. and Al-Saqur, I. M. (1980). The prevalence of hydatid cysts in slaughtered animals in Iraq. Annals of Tropical Medicine and Parasitology, 74:185-187.

Al-Saqur, I. M.; Fahelbab, G. O.; Awath, F.; Abdulkader, R. M. and Imdi, K. A. (2001). Preliminary studies in the epidemiology of Hydatid cysts in Sebha Great Jamahirya. Western Medical Veterinary conference, 18th, Tripoli – Libya.

Altinatas, N. and Ozensoy, S. (1993). Hydatidosis in Turkey.

International Archives of Hydatidology, 31:98.

Al-Yaman, F. M.; Assaf, L.; Hailat, N. and Abdel-Hafez, S. K. (1985). Prevalence of hydatidosis in slaughtered animals from North Jordan. *Annals of Tropical Medicine and Parasitology*, 79: 501 – 506.

Anon. (1993). The surgical incidence rate of hydatidosis in Tunisia (1988 – 1992). Report of the S.S.S.B (Direction de la Santé et des Soins de Base). Ministry of public Health, Tunis.

Awan, M. A. Q.; Gusbi, A. M. and Beesley, W. N. (1990). Echinococcus in Libya. III. Further studies of *Echinococcus granulosus* in dogs. *Annals of Tropical Medicine and Parasitology*, 84:473-475.

Babars, M. A.; Fayad, M. F. and Arafa, M. A. (1987). The incidence of hydatidosis among populations in Sucz Canal zone. *Journal of the Egyptian Society Parasitology*, 17 (2): 377 – 380.

Behir, A.; Jemni, L.; Allegue, M.; Hamdi, A.; Khalifa, K.; Letaif, R.; Mlika, N.; Dridi, H.; Larouze, B.; Rousett, J. J.; Gaudebout, C. and Jemmali, M. (1985). Epidemiologie de l'hydatidose dans le Sahel et le Centre Tunisiens. Bulletin de la Société Pathologie Exotique, 78:687-690.

Behir, A.; Larouze, B.; Soltani, M.; Hamdi, A.; Bouhaouala, H.; Ducic, S.; Bouden, L.; Ganouni, A.; Achour, H.; Gaudebout, C.; Rousset, J. and Jemmali, M. (1991). Echotomographic and serological

population-based study of hydatidosis in central Tunisia. *Acta Tropica*, 49:149-153.

Ben-Osman , F. (1965) . Considerations epidemiologiques sur, L'hydatidose animale en Tunise . Archives de l'Instit Pasteur . *Tunis* ,3-4 : 409-418 .

Bouree, P. (2001). Hydatidosis: dynamics of transmission. World

Journal Surgery, 25 (1): 4-9.

Bowles, J. Blair, D. and McManus, D.P. (1992). Genetic Variants within the genus *Echinococcus* identified by DNA mitochondrial sequencing *Molecular Biochemical and Parasitology*, 54: 165-174.

Bowles, J. and McManus, D.P. (1993a). Molecular Variation in Echinococcus, Acta Tropica, 53: 291 – 305.

Bowles, J. and McManus, D.P. (1993b). Rapid discrimination of *Echinococcus* species and strains using aPCR- based RFLP method. *Molecular Biochemistry and Parasitology*, 57:231-239.

Braddick, M. R. and Reily, W. J. (1993). Human hydatid disease leading to hospital admission in Scotland 1968 – 1989. *Health Bulletin*, 7:80-85.

Castigliola, L. (1918). Un caso di echinococcosis epatica en cirenica (Libya) Giornal-medica Millan, 66: 1096 – 1099.

Cicogna , D. (1961) . L'echinococcosis in Tripolitania . Bolletino Sanitario della Tripolitania , 2 : 3 .

Clavel, A.; Varea, M.; Doiz, O.; Lopez, L.; Quilez, J.m Cqsti-Ilo, F. J.; Rubio, C. and Gomez-Lus, R. (1999). Visualization of hydatid elements: comparison of several techniques. *Journal Clinical Microbiology*, 37 (5): 1561 – 1563.

Craig, P. S.; Deshan, L. and Zhaoxun, D. (1991). Hydatid disease in China. *Parasitology Today*, 7:46 50.

Craig, P. S.; Gasser, R. B.; Parada, L.; Cabrera, P.; Parietti, S.; Borgues, C.; Acuttis, A.; Agulla, J.; Snowden, K. and Paolillo, E. (1995). Diagnosis of canine echinococcosis comparison of copoantigen and serum antibody tests with arecoline purgation in Uruguay.

Veterinary Parasitology, 56: 293-301.

Constantine, C. C.; Thompson, R. C. A.; Jenkins, D. L.; Hobbs, R. P. and Lymbery, A. J. (1993). Morphological characterization of adult *Echinococcus granulosus* as a mean of determining transmission patterns.

Journal of Parasitology, 79:57-61.

Cousi, D. (1951). L'échinococcose en Tunisie. Afrique Française Chirurgicale, 4:379-386.

Dada, B. J.; Adegboye, D. S. and Mohamed, A. N. (1980). The epidemiology of Echinococcosis infection in Kano state, Nigeria. *Annals of Tropical Medicine and Parasitology*, 74 (5): 515-517.

Daily, M. D. and Sweatman, G. K. (1965). The taxonomy of Echinococcus granulosus in the dromedary in Lebanon and Syria.

Annals of Tropical Medicine and Parasitology, 57: 463-477.

Dajani, Y. F. (1978). Prevalence of hydatid disease in Syria and Jordan preliminary results. Transactions of the Royal Society of Tropical Medicine and Hygiene, 72:320-321.

Dajani, V. F. and Khalaf, F. H. (1981). Hydatidosis and tenuicollosis in sheep and goats of Jordan: a comparative study. *Annals of Tropical Medicine and Parasitology*, 75: 175 – 179.

Dar, F. K. and Taguri, S. (1978). Human hydatid disease in Eastern Libya. Transactions of the Royal Society of Tropical Medicine and Hygiene, 72:313-314.

Dar, F. K. and Taguri, S. (1979). Epidemiology and epizoology of hydatidosis in the Libyan Jamahiriya and recommendations for a programme of surveillance and control of the disease. *Garyounis Medical Journal*, 2:11-15.

Develoux, M.; Lamothe, E.; Sako, A.; Landois, J.; Straboni, J. P. and Vetter, J. M. (1985). Deux cas d'hydatidose en Republique du Niger. Bulletin de la Societe de Pathologie Exotique et de ses Filiales, 78: 216-220.

Devendra, C. and Mcleroy, G. B. (1982). Goats and sheep production in the Tropics. (eds. W. J. A. Payne). Longman, London pp:120-132.

Dyab, K. A.; Hassanein, R.; Hussein, A. A.; Metwally, S. E.; and Gaad, H. M. (2005). Hydatidosis among man and animals in Assiut and Aswan Governorates. *Journal of the Egyptian Society of Parasitology*. 35 (1): 157-166.

Eckert, J. and Thompson, R. C. A. (1988). Echinococcus strains in Europe: a review. Tropical Medicine and Parasitology, 39:1-9.

El-Boulaqi, H. A. and Taguri, S. (1980). Incidence of human hydatid disease in Eastern Libya. *Journal of the Egyptian Society of Parasitology*, 10 (1): 101 – 106.

El-Kordy, M. I. (1946). On the incidence of hydatid disease in domestic animals in Egypt. *Journal of the Egyptian Medical Association*, 29:267-279.

Et-Sageyer, M. M. (1989) Morphological, biochemical characterization and serological tests of larval cysts of *Echinococcus granulosus* in human and sheep hosts. A thesis presented to the department of zoological science, in partial fulfillment for the requirements for the degree of Master of Science in zoology.

El-Sageyer, M. M. and Kidwai, S. A. (1993). Observations on morphology and viability of protoscoleces from larva of *Echinococcus granulosus* in Libyan Arab Jamahiriya. *Rivista di Parasitologia*, X: 389-398.

Ensminger, M. E. (1983). Animal Science. 8th, ed. The interstate printeis & publishes, Inc. Danville, Illinois pp: 88 - 96.

Erman, T.; Tuna, M.; Göcer, I.; Ildan, F.; Zeren, M. and Cetinalp, E. (2001). Intracranial intraosseous hydatid cysts. *Neuroswy Focus*, 11 (1): 1-3.

Euzéhy, J. (1991). The epidemiology of hydatidosis with special reference to the Mediterranean area. *Parassitologia*, 33:25-39.

FAO (1993). Zoonotic disease in the Near East Region. Regional Office for the Near East, Cairo.

Fossati, G. J. (1970). Las parasitosis respiratorios halladas en pacientas arabolibicos de cirenaica (Libya) en Los ulumos diez anos. II. Hidatidosis toracica. *Revista Iberica de parassitologia*, 30:587 – 647.

Gabriele, F.; Bortletti, G.; conchedda, M. and Palmas, C. (1997). Epidemiology of hydatid disease in the Mediterranean basin.

Parasitology, 39:47-52.

Gdourra, N. M. (2003). Prevalence, Morphological and Biochemical Studies of the metacestode of *Echinococcus granulosus* in Camels (*Camelus dromedaries*) in Sirt, Libya. A thesis submitted in partial fulfillment and have of the requirements of MSc in zoology.

Gebreel, A. O.; Gilles, H. M. and Prescott, J. E. (1983). Studies on the sero-epidemiology of endemic diseases in Libya. I. Echinococcosis in Libya. *Annals of Tropical Medicine and Parasitology*, 77:391 – 397.

Gemmell, M. A. and Lawson, J. R. (1986). Epidemiology and control of hydatid disease. In: Thompson, R. C. A. (ed.) the Biology of

Echinococcus and hydatid disease. George Allen and Unwin, London, pp. 189-216.

Gharbi, H. A.; Hassine, W. and Abdesselom, K. (1985). L'hydatidose abdominale à l'échographie. Reflexions et aspects particuliers Echinococcus granulosus. Annals of Radiology, 28:31-34.

Gottstein, B. (2000). Epidemiology and systematic of cystic and alveolar hydatid disease. Chirrurg, 71 (1): 1-8.

Gracey, J. F. and Collins, D. S. (1992). Meat Hygiene, Bailliere Tindall, London pp: 56 - 82.

Gusbi, A. M. (1987). Echinococcosis in Libya. I. Prevalence of Echinococcus granulosus in dogs with particular reference to the role of dog in Libyan society. Annals of Tropical Medicine and Parasitology. 81 (1): 29 – 34.

Gusbi, A. M.; Awan, M. A. Q. and Beesley, W. N. (1987). Prevalence of hydatidosis (Echinococcus granulosus) in Sheep. Annals of Tropical Medicine and Parasitology, 81:35-41.

Gushi, A. M.; Awan, M. A. Q. and Beesley, W. N. (1990). Echinococcosis in Libya. IV. Prevalence of hydatidosis (Echinococcus granulosus) in goats, cattle and camels. Annals of Tropical Medicine and Parasitology, 84:477-482.

Gushi, A. M.; Awan, M. A. Q. and Beesley, W. N. (1991). Experimental infection of Libyan sheep with Echinococcus granulosus.

Annals of Tropical Medicine and Parasitology, 85:433-437.

Gutierrez, Y. (1990). Diagnostic Pathology of Parasitic Infections with Clinical Correlations. Lea & Febiger, Philadelphia.

Hamdy, I. I.; Mikhail, E. A.; Soliman, A. A. and Hamed, H. H. (1980). A study on hydatidosis in some animals in Egypt. *Journal of the Egyptian Society of Parasitology*, 10 (1): 43 – 51.

Haridy, F. M.; Ibrahem, B. B. and Morsy, T. A. (1998). Studies on hydatidosis in slaughtered camels in Egypt. Journal of the Egyptian Society of Parasitology. 28 (3): 673-681.

Haridy, F. M.; Badaweia, B.; Ibrahem and Tosson, A. M. (2000). Sheep-dog-man. The risk of zoonotic cycle in hydatidosis. *Journal of the Egyptian Society of Parasitology*, 30 (2): 423-429.

Harris, A.; Heath, D. D.; Lawrence, S. B. and Shaw, R. J. (1989). Echinococcus granulosus: ultrastructure of epithelial changes during the first 8 days of metacestode development in vitro. International Journal for Parasitology, 19:621-629.

Hegazi, M. M.; Abdel-Magied, S. A.; Abdel-Wahab, F. M. and Atia,
R. A. (1986). Epidemiological study of echinococcosis in 1 Dakahlia governorate, Egypt. Journal of the Egyption Society of Parasitology,
16:541-548.

Himonas, C.; Antoniadou-Sotiriadou, K. and Papadopoulos, E. (1994). Hydatidosis of food animals in Greece: Prevalence of cysts containing viable Protoscoleces. *Journal of Helminthology*, 68:311-313.

Hobbs , R. P.; Lymbery , A. J. and Thompson , R. C. A. (1990) . Rosterllar hooks morphology of *Echinococcus granulosus* (Batsch, 1786) from natural and experimental Australian hosts , and its implication for . strain recognition . Parasitology , 101 : 274 – 281 .

Ibrahem, M. M. and Craig, P. S. (1998). Prevalence of cystic echinococcosis in camels (Camelus dromedaries.) in Libya. Journal of Helminthology, 72:27-31.

Ibrahem, M. M. and Gusbi, A. M. (1997). Cystic echinococcosis in North Africa (excluding Morocco): Veterinary aspects In: Provo, Utah, USA. Compendium on Cystic Echinococcosis in Africa and Middle Eastern Countries with Special Rreferences to Morocco, (eds. F.L. Andersen, H., Ouhelli and Kachani, M) Brigham Young University, Provo, Utah, USA.

Irshadullah, M.; Nizami, W. A. and Macpherson, C. N. L. (1989). Observation on the suitability and importance of the domestic intermediate hosts of *Echinococcus granulosus* in Uttah Pradesh, India. *Journal of Helminthology*, 63:39-45.

Islam, A. W. M. S. (1982). The prevalence of hydatid cysts in slaughtered cattle in Bangladesh. *Journal of Helminthology*, 56: 247-250.

Jaiem , A. (1984) . L'échinococcose hydatique dans la région de Sousse : enquête épidémiologique . *Maghreb Veterinarian* , 1:9-12 .

Kachani , M.; Ouhelli , H.; El-Hasnaoui , M. and Andersen , F. L.
(1998) . Environmental adaptation of Echinococcus granulosus in Morocco . Symposium on environmental adaptation of Echinococcosus .
18 - 20 August , 1998 , Hokkaido University Conference Hall Centennial Hall .

Kalani, B. P.; Ojha, S. N., Bhargava, R. K. and Broadhead, R. L. (1984). Hydatid disease in children in Libya. *Annals of Tropical* Pediatrics, 4: 195 – 199.

Kamhawi, S. and Hijjawi, N. (1992). Current studies on the epidemiology of unilocular hydatidosis in Jordan and its social implications. *Report on Parasitic Diseases of the Middle East*. National Institutes of Health, Bethesda, Maryland, p. 8.

Karpathios, T.; Fretzayas, A.; Nicolaidou, P.; Papadellis, F.;
Vassalos, M.; Tselentis, J.; Thomaidis, T. and Matsaniotis, N.
(1985). Statistical aspects of hydatid disease in Greek adults. American Journal of Tropical Medicine and Hygiene, 34:124-128.

Khan, A. H. and El-Buni, A. A. (1999). Prevalence of hydatid cystic in slaughtered animals in Benghazi, Libya. Sebha Medical Journal, 2 (1):7-9.

Khan, A. H.; El-Buni, A. A. and Ali, M. Y. (2001). Fertility of the cysts of *Echinococcus granulosus* in domestic herbivores from Benghazi,

Libya and the reactivity of antigens produced from them. Annals of Tropical Medicine & Parasitology, 4:337-342.

Khan, A. H.; El-Sageyer, M. M. and Kdwai, S. A. (1990). Seroepidemiology study of human hydatid disease in Libya using finger prick blood samples. *Annals of Tropical Medicine and Parasitology*, 84:537-539.

Khan, A. H. and Kidwai, S. A. (1996). Management of immunodiagnostic tests in human hydatidosis in Libya. *Rivista di Parassitologia*, XIII: 473 – 477.

Krammerer, W. S. and Schantz, P. M. (1993). Echinococcal disease.

Infectious Disease Clinics of North America, 7:605-618.

Laajan, M. A. and Nouh, M. S. (1991). Hydatidosis: clinical significance and morbidity patterns in Saudi Arabia. East African Medical Journal, 68:57-63.

Lahmar, S.; Kilani, M.; Torgerson, P. R. and Gemmell, M. A. (1999). Echinococcus granulosus larvae in the livers of sheep in Tunisia: the effects of host age. Annals of Tropical Medicine and Parasitology, 93:75-81.

Larbaoui, D. and Allyulya, R. (1979). Etude épidémiologique de l'hydatidose en Algérie. Résultats de deux enquêtes rétrospectives portant sur 10 ans. *Tunisie Médécine*, 57:318326.

Larbaoui, D.; Allyulya, R.; Osiiskaya, L. V.; Yu Osiiskii, I. X. and Benel'Muffok, M. (1980). Hydatidosis in Algeria *Meditsinskaya* Parasitarnye Bolezni, 49:21-28.

Lavikainen, A.; Lehtinen, M. J.; Meri, T.; Hirvela-Koski, V. and Meri, S. (2003). Molecular genetic characterization of the Fennoscandian Cervid strain a new genotypic group (G10) of Echinococcus granulosus. Parasitology, 127: 207-215.

Löscher, T.; Von Sonnenburg, F. and Nothdurft, H. D. (1992). Epidemiology and therapy of human echinococcosis in Central Europe. 8th International Congress for Tropical Medicine and Malaria. Pattaya, Thailand, Vol. 1 p. 342.

Macchioni, G.; Lanfranchi, P.; Abdallatif, M. A. and Testi, F. (1985). Echinococcosis and hydatidosis in Somalia. Bulletin of Science of the Faculty of Zoology and Veterinary Medicine, 5:179-189.

Macpherson, C. N. L. (1981). Epidemiology and strain differentiation of *Echinococcus granulosus* in Kenya. PhD thesis. University of London.

Macpherson, C. N. L. (1985). Epidemiology of hydatid disease in Kenya: a study of the domestic intermediate hosts in Masailand.

Transactions of the Royal Society of Tropical Medicine and Hygiene, 79: 209 – 217.

Macpherson, C. N. L.; Craig, P. S.; Romig, T.; Zeyhle, E. and Watschinger, H. (1989). Observations on human echinococcosis

(hydatidosis) and evaluation of transmission factors in the Maasai of northern Tanzania. *Annals of Tropical Medicine and Parasitology*, 84:489-497.

Macpherson, C. N. L.; Wachira, T. M.; Zeyhle, E.; Roming, T. and Macpherson, C. (1986). Hydatid disease: Research and control in Turkana, IV. The pilot control program. Transactions of the Royal Society of Tropical Medicine and Hygiene, 80: 196-200.

Magnussen, P. and Thorsen, S. (1992). Anaphylactic shock caused by rupture of hydatid cysts. *Ugeskr Laeger*, 151:1125-1126.

Mahdi, N. K. and Benyan, A. K. (1990). Hydatidosis among Iraq children. Annals of Tropical Medicine and Parasitology, 84:289-292.

Matossian, R. M.; Rickard, M. D. and Smyth, J. D. (1977). Hydatidosis a global problem of increasing importance. Bulletin of the World Organization, 55: 499 - 507.

Mazyed, M. A. M.; Mostafa, M. M. and Morsy, T. A. (1998). Spinal cord hydatid cyst in Egypt. Journal of Egyptian Society Parasitology, 28 (3): 665 - 668.

McManus, D. P. (1981). A biochemical study of adult and cystic stages of Echinococcus granulosus of human and animal origin from Kenya. Journal of Helminthology, 55:21-27.

McManus, D. P. and Bryant, C. (1995). Biochemistry, Physiology and Molecular Biology of Echinococcus. In: Echinococcus and hydatid

disease (eds., R.C.A Thompson and A. J. Lymbery). CAB International, Wallington. U.K., pp: 135 - 171.

Medulla, C. (1931). La cirenaica del punto di vista sanitario. Archivio Italiano di Scienze Mediche Coloniale, 12:391-418.

Mohamed, S. A.; Abdel-Hasseeb, S. M. and Bayomi, A. H. (1997). Incidence and pathology of lung hydatidosis in slaughtered *Camelus dromeduries* at Assiut Governorate. *Beni-Suef Veterinary Medical Research*, 7 (2): 173 – 187.

Mohamed, A. A.; Fadiel, M. M. and Fateh-Elbab G. A. (2004). Prevalence and epidemiology of hydatidosis in Local and imported animals Sebha City. *Journal of Sebha University*, vol. 3 (3): 39-51.

Morris, D. L. and Richards, K. S. (1992). Hydatid Disease. Current medical and surgical management. Butterworth-Heinemann Ltd., Oxford.

Muller, R. (2002). Worms and human disease, second edition, CAB International Publishing, printed and bound in the U.K. by Biddles Ltd., Guildford and King's Lynn.

Munzer, D. (1991). New perspectives in the diagnosis of Echinococcus disease. Journal of Clinical Gastroenterology, 13:415-423.

Navarrete, I.; Serrano, F.; Perez, E.; Brena, M.; Morales, I. and Gil, F. (1991). Study of prevalence of canine echinococcosis in Extremadura: possible influence over ovine production. Archivos de la Hidatidosis, 30: 1253 – 1260.

- Nourian, A. A.; Ataeian, A. and Hanilou, A. (1997). Hydatidosis / echinococcosis in Zanjan area, north-west of Iran. XVIII International Congress of Hydatidology, Lisbon.
- Onah, D. N.; Chiejna, S. N. and Emehelu, C. O. (1989). Epidemiology of echinococcosis / hydatidosis in Anambra state, Nigeria.

 Annals of Tropical Medicine and Parasitology, 83 (4): 387 393.
- Onorato, R. (1927). Echinococcosis in Tripoli Tania. Archivio Italiano ds Scienze Mediche, 8, 9.
- Ouhelli, H. and Dakkak, A. (1992). Hydatidosis in Morocco. Report on Parasitic Diseases of the Middle East. National Institutes of Health, Bethesda, Maryland, p. 1-2.
- Packer, D. E. and Ali, T. M. (1986). Echinococcus granulosus in dogs in Libya. Annals of Tropical Medicine and Parasitology, 80: 137-139.
- Pandey, V. S.; Dakkak, A. and Elmamoune, M. (1987). Parasites of stray dogs in the Rabat region, Morocco. Annals of Tropical Medicine and Parasitology, 81:53.
- Pandey, V. S.; Ouhelli, H. and Ouchtou, M. (1986). Hydatidosis in sheep, goats, and dromedaries in Morocco. *Annals of Tropical Medicine and Parasitology*, 80:525-529.

Pandey, V. S.; Ouhelli, H. and Moumen, A. (1988). Epidemiology of hydatidosis / Echinococcosis in Ouarzazate, the pre-Saharian region of Morocco. *Annals of Tropical Medicine and Parasitology*, 82: 461 – 470.

Papadopoulos, G. (1981). Conclusion and recommendations on Libya mission, draft report.

Rahman, M. S. (1982). Dystocia due to hydatid disease of the pelvis. Helminthological Abstracts, 51 (12).

Rausch, R. L. (1995). Life Cycle Patterns and Geographic Distribution of *Echinococcus* Species. In: *Echinococcus* and Hydatid Disease. (eds. R. C. A. Thompson and A. J. Lymbery). CAB International. Wallington, U. K.

Rausch, R. L. (1997). Echinococcus granulosus: Biology and Ecology. In: Compendium on Cystic Echinococcosis in Africa and In Middle Eastern Countries with Special Reference to Morocco. (eds. F. L. Andersen, H. Ouheli and M. Kachani). Brigham Young University Provo, Utah, USA.

Roberts, M. G.; Lawson, J. R. and Gemmell, M. A. (1986). Population dynamics in echinococcosis and cysticercosis: mathematical model of the life-cycle of *Echinococcus granulosus Parasitology*, 92: 621-641.

Romia, S. A.; Youssef, M. E.; Handousa, A. E.; Rizk, H. and Sallam, S. M. (1992). Dot-ELISA as a diagnostic test in hydatid diseases. *Journal of Egyptian Society Parasitology*, 22 (3): 603 – 610.

- Saad , M. B. and Fatehell-Bab , O. G. (1996) . A study on echinococcosis in slaughtered camels in Sebha . Presented in the first Scientific meeting of Libyan Society of Veterinary Medical Sciences , Tripoli , 2-3.
- Saad, M. C. and Magzoub, M. (1986). Echinococcus granulosus infection in dogs in Tambool, Sudan. Journal of Helminthology, 60: 299-300.
- Saad, M. B.; Zien-Eldin, E. A. and Tag-Eldin, M. H. (1983). Some observations on the prevalence and pathology of hydatidosis in Sudanese camels (Camelus drumedarus). Revue d'Elevage et de Medicine Veteriane des Pays Tropicaux, 36:359 363.
- Saeed, I.; Kapel, C.; Saida, L. A.; Willingahm, L. and Nasen (2000). Epidemiology of *Echinococcus granulosus* in Arbil province, northern Iraq, 1990 1998. *Journal of Helminthology*, 74-83 88.
- Saidi, F. (1976). Surgery of Hydatid Disease. W. B. Saunders Company Ltd., London.
- Schaefer, J. W. and Khan, M. Y. (1991). Echinococcosis (hydatid disease): lessons from experience with 59 patients. *Reviews of Infectious Dideases*, 13: 243 247.
- Schantz, P. M.; Chai, J.; Craig, P. S.; Echert, J.; Jenkins, D. L.; Macpherson, C. N. L. and Thakur, A. (1995). Epidemiology and Control of Hydatid Disease. In: *Echinococcus* and Hydatid Disease.

(eds. R. C. A. Thompson and A. J. Lymbery). CAB International, Wallington, U. K.

Schwabe, C. W. (1968). Epidemiology of Echinococcosis. Bulletin of the World Health Organization, 39: 131 - 135.

Schwabe, C. W. and Abou Daoud, K. (1961). Epidemiology of echinococcosis in the Middle East. I. Human infection in Lebanon, 1949-1959. American Journal of Tropical Medicine, 10:374-381.

Senevet, G. (1951). Epidemiologe du kyste hydatique en Afrique du nord. Archivos Internacionales de Hidatidologia, 12:113-120.

Shaafie, C. A.; Khan, A. H. and Rambabu, K. (1999). Biochemical profiles of hydatid cyst fluids of *Echinococcus granulosus* of human and animal oringin in Libya. *Journal of Helminthology*, 73: 255 – 258.

Shambesh, M. K. A. (1997). Human cystic echinococcosis in North Africa (excluding Morocco). In: Compendium on Cystic Echinococcosis in Africa and Middle Eastern Countries with Special References to Morocco, (eds. F.L. Andersen, H., Ouhelli and Kachani, M) Brigham Young University, Provo, Utah, USA.

Shambesh, M. A.; Craig, P. S., Macpherson, C. N., Rogan, M. T., Gusbi, A. M. and Echtuish, E. F. (1999). An extensive ultrasound and serologic study to investigate the prevalence of human cystic echinococcosis in northern Libya. *Journal of Tropical Medicine and Hygiene*, 60:462-468.

Shambesh, M. K.; Macpherson, C. N. L., Beesley, W. N., Gusbi, A. and Elsonosi, T. (1992). Prevalence of human hydatid disease in northwestern Libya: a cross-sectional ultrasound study. *Annals of Tropical Medicine and Parasitology*, 86:381-386.

Shweiki, H. M.; Hira, P. R. and Behbehani, K. (1990). Cystic hydatid disease: aspects of the incidence in man in Kuwait, Arabian Gulf. European Journal of Epidemiology, 6:15-19.

Sirol, J. and Lefevre, M. (1971). L'hydatidose humaine au Tchad pays d'élévage. A propos de trois observations colligées à Fort Lamy, Chad. Bulletin de la Société de Pathologie Exotique, 64:887-889.

Smyth, J. D. and Davies, Z. (1974). Occurrence of physiological strain of *Echinococcus granulosus*, demonstrated by in vitro culture of protoscoleces from sheep and horse hydatid cyst. *International Journal for Parasitology* 4:442-445.

Sultan Sheriff, D.; El-Fakhri, M. and Kidwai, S. A. (1989). Lipids in hydatid fluids collected from lungs and liver of sheep and man. Journal of Helminthology, 63:266-268.

Sweatman, G. K. and Williams, R. J. (1963). Comparative studies on the biology and morphology of *Echinococcus granulosus* from domestic livestock, moose and reindeer. *Parasitology*, 53:339-390.

Tashani, O. A.; Zhang, L. H., Boufana, B. Jegi, A. and McManus D. P. (2002). Epidemiology and strain characteristics of *Echinococcus*

granulosus in the Benghazi area of Eastern Libya. Annals of Tropical Medicine and Parasitology, 96(3):1-12.

Thompson, R. C. A. (1995). Biology and Systematics of *Echinococcus*.
In: *Echinococcus* and Hydatid Disease. (eds. R. C. A. Thompson and A. J. Lymbery) CAB International, Wallington, U.K.

Thompson, R. C. A. and Lymbery, A. J. (1988). The nature, extent and significance of variation within the genus *Echinococcus*. Advances in Parasitology, 27:209-258.

Thompson, R. C. A. and Lymbery, A. J. (1990). Echinococcus: biology and strain variation. International Journal for Parasitology, 20: 457-470.

Thompson, R. C. A. and Lymbery, A. J. (1991). The epidemiological significance of biological variation in *Echinococcus . Archivos de la Hidatidosis*, 30:195-200.

Thompson, R. C. A.; Lymbery, A. J. and Constantine, C. C. (1995). Variation in Echinococcosis: towards a taxonomic revision of the genus.

Advances in Parasitology, 35: 145 – 176.

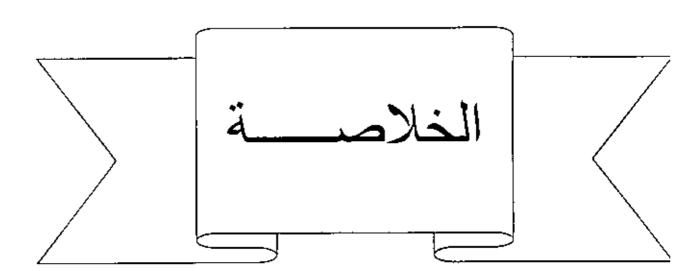
Thompson, R. C. A. and Mc Manus, D. P. (2002). Towards a taxonomic revision of the genus Echinococcus. *TRENDS* in *Parasitology*, 10:452-457.

Watson-Jones, D. L.; Criag, P. S.; Badamochir, D.; Rogan, M. T.; Wen, H.; Hind, B. (1997). A pilot, serological survey for cystic echinococcosis in North-Western Mongolia. *Annals of Tropical Medicine and Parasitology*, 9 (2): 173-177.

Williams, J. F.; Adaros, H. L. and Trejos, A. (1971). Current prevalence and distribution of hydatidosis, with special reference to the Americas. *Annals of Tropical Medicine and Parasitology*, 20:224-235.

Wilson, J. E. and Rausch, R. L. (1980). Alveolar hydatid disease. A review of clinical features of 33 indigenous cases of *Echinococcus multilocularis* infection in Alaskan Eskimos. *American Journal of Tropical Medicine and Hygiene*, 29:134-155.

Wosene, A. (1991). Traditional husbandry practices and major health problems of camels in the Ogaden (Ethiopia). *Nomadic Peoples*, 29:21-30.



الخلاص____ة

تم فحص 8123 من الأغنام و 3794 من الماعز و 113 من الأبقار و 739 من الإبل خلال الفترة مابين يوليو 2004 وحتى مايو 2005 ، والتي ذبحت في السلخانة المعتمدة في مدينة سرت. وتبين من الدراسة أن نسبة الإصابة بالأكياس العذارية (الطور اليرقى للمشوكة الحبيبية) في هذة الحيوانات قد بلغت 4.9 % في الأغنام ، 2.4 % في الماعز و 15 % في الأبقار و 2.7 % في الإبل.

أوضحت الدراسة أن الإصابة بالأكياس العذارية قد سجلت في كلا الجنسين الذكور والإناث لجميع أنواع الحيواتات المفحوصة وتبين من الدراسة أنه لا تأثير لجنس الحيوان على الإصابة في الأغنام والماعز والأبقار ولكن توجد اختلافات معنوية قليلة في الإبل. كما أوضحت الدراسة أن انتشار الإصابة بالأكياس العذارية يزداد بزيادة العمر. ونتيجة للنمو البطئ للأكياس العذارية في الإعمار الصعيرة من الحيوان.

أظهرت الدراسة عدم وجود فروق معنوية في مدى انتشار الإصبابة بالأكياس العذاريه في الأغنام والماعز والإبل المفحوصة على مدار السنة بالنسبة لنوع الحيوان الواحد

تم تحديد مكان الإصابة بالأكياس العذارية للحيوانات المفحوصة حيث استأثر الكبد في الأغنام بنسبة 77.9 بينما الرنتين 48.9 أما الماعز فإصابة الكبد كانت 77.9 والرنتين 60 % ونسبة إصابة الكبد في الأبقار 64.7 % وفي الرنتين 68 % ، أما الإبل فالرنتين كانت أكثر إصابة بنسبة 75 % عن تلك في الكبد 65 % . أظهرت الدراسة أنه لا تأثير لجنس الحيوان على مكان الإصابة بالأكياس العذارية في الأغنام والأبقار والإبل بينما في الماعز تبين وجود اختلافات معنوية بين الجنسين ومكان الإصابة (10.00 = 9) . كما أظهرت الدراسة أن للعمر تأثير على مكان الإصابة في الأغنام والماعز (10.00 = 9) ولكن هذا التأثير لم يسجل في الأبقار والإبل .

شدة الإصابة في الأغنام تتراوح بين 1 - 20 كيس في كلاً من الكبد والرنتين ، وأظهرت النتائج أن 53.8 % إصابة متوسطة (4 - 8 الكياس) و 36 % إصابة متوسطة (4 - 8 الكياس) و 36 % إصابة متوسطة (4 - 8 الكياس) و 30.0 % إصابة كثيفة (8 - 20 كيس) . وفي الماعز 61.6 % الإصابة بالأكياس

خفيغة و 28.7 % متوسطة و 9.7 % كثيغة , بينما الأبقار شدة الإصبابة تمثلت بنسية 52.9 % خفيفة و 35.3 % متوسطة و 11.8 % كثيفة , أما الإبل فشدة الإصبابة بها 40 % خفيفة و 35 % متوسطة و 25 % كثيفة .

أظهرت النتائج وجود فروق معنوية بين شدة الإصابة والعمر في الأغنام والماعز (P = 0.000) ، ولم توجد هذة الغروق في حالمة الأبقار (P = 0.699) والإبل (P = 0.053) . أما جنس الحيوان فلم يكن لمه تأثير على شدة الإصابة بالأكياس العذاريه .

أكثر الأعضاء إصابة بالأكباس العذارية في الأغنام هي الكبد، وأظهر التقصي نسبة كبيرة جداً للأكباس ذات الحجم المتوسط (2 – 5 سم) بنسبة 56.7 %، بينما الرئتين كانت مصابة بالأكباس صغيرة الحجم (> 2 سم) بنسبة 45.1 %، نفس أحجام الأكباس سجلت في الماعز بنسبة 45.8 % من الحجم المتوسط (2 – 5 سم) في الكبد وبنسبة 42.8 % في الرئتين من الحجم الصغير، وأظهرت الدراسة أن معظم الإصابة في الأبقار والإبل كانت بالأكباس ذات الحجم الصغير (> 2 سم) في كل من الكبد والرئتين، وتبين من النتائج أن الأكباس ذات الحجم الحبغير (> 2 سم) في كل من الكبد والرئتين، وتبين من النتائج أن الأكباس ذات الحجم الكبير (5 – 10 سم) سجلت فقط في الماعز.

أظهرت الدراسة أن نسبة الخصوبة في الأكياس العذارية تساوي 72 % في الأغنام و 58 % في الأغنام و 58 % في الأبل , أما جميع الأكياس العذارية في الأبقار كانت عقيمة , وتبين من الدراسة أنه لا تأثير لعمر الحيوان أو الجنس على خصوبة الأكياس العذارية في أي نوع من الحيوانات المفحوصة (P > 0.05) .

أشارت الدراسة إلى أن حبوية الأكياس العذارية في الأغنام والإبل كانت بنسبة 70% في كلاهما وهي أكثر نسبة من حبوية الأكياس في الماعز (58%). تبين من الدراسة أن حبوية الأكياس العذارية كانت بنسبة 80% و 76% في أكباد الأغنام والإبل على التوالي وهي أكثر حبوية من تلك الموجودة في الرئتين 55% في الأغنام و 62% في الإبل. بينما حبوية الأكياس العذارية في الماعز كانت 66% في الرئتين و 50% في الكبد. أوضنحت الدراسة عدم وجود فروق معنوية بين حبوية الأكياس العذارية في الكبد والرئتين للحبوانات المفحوصة (P=0.01) فروق معنوية بين حبوية الأكياس العذارية في الكبد والمرئتين وجود اختلافات معنوية بين حبوية الأكياس العذارية وحجمها في جميع الحبوانات قبد

أظهرت الدراسة الشكلية (المورفولوجيه) لرؤوس الديدان في الأكباس المائية عدم وجود فروق كبيرة في الحجم في حالة الأغنام والماعز والأبقار ، أما في الإبل تبين أن الرؤوس كانت أكبر حجما , ومن خلال الدراسة المورفولوجيه للخطاطيف اظهرت عدم وجود اختلاف في العدد في الكند والرئتين لكل نوع من الحيوانات ، كما تبين عدم وجود أي فروق معنوية بين حجم الخطاطيف ونوع الحيوان .

أوضحت النتائج المتحصل عليها من هذة الدراسة إلى أن الأغنام هي أكثر الحيوانات، يليها الماعز فالإبل، ثم الأبقار ورغم أن الأبقار هي الأقل ذبحا إلا أنها أكثر إصابة وذلك ربما يعزى إلى أنها في أعمار متقدمة وتكون قد تعرضت للإصابة ببيوض الديدان المشوكة الحبيبية أكثر فترة. كما أوضحت الدراسة أن الماعز هي أقل الحيوانات إصابة رغم أنها ترعي مع الأغنام في نفس الأماكن وريما يفسر ذلك باختلاف طريقة التعنية بين النوعين، حيث أن الماعز تتعذي على الشجيرات والنباقات العالية نسبيا التي غالبا ما تكون ملوثه ببيوض المشوكة الحبيبية أما الحيوانات الأخرى كالأغنام فأنها تتعذي على الحشائش القصيرة حيث كثيرا ما تنتشر بيوض الدودة.

أشار كثير من البحاث إلى أن الإبل تلعب دوراً مهما في استمرار دورة حياة المشوكة الحبيبية نتيجة لأرتفاع نسبة إصابتها بالأكباس المانية إلا أن أعداد الإبل لا تقارن باعداد الأغذام التي تساهم في استمرار دورة حياة الطفيل لعدة أسباب منها ، عدد الأغذام الكبير الذي قد بصل إلى أكثر من 8 ملايين ، بينما لا يتجاوز عدد الإبل نصف مليون ، وتذبح الأغنام بأعداد اكبر من الإبل ، كما أن نسبة خصوبة الأكباس المانية في الأغنام أكثر من تلك في الإبل .

النتائج المتحصل عليها في هذة الدراسة تشير إلى أهمية الدراسات التقصيلية للأكياس المانية من حيث شدة الإصابة والخصوبة وتشير الدراسة إلى أن الكبد أكثر إصابة من المرنتين وحيوية الأكياس في الكبد أكثر من تلك في الرنتين ، ولذلك تعتبر الكبد عضو مهم في استمرار دورة حياة الدودة ، وحيث أن الكبد يعتبر هو العضو المفضل لدى المستهلك الليبي ويميل القصابون إلى عدم المتصحية بالكبد المصابة بأكملها ويتم فصل الجزء المصاب فقط الذي يتم التخلص منه بطريقة غير سايمة و غالباً ما تلتهمه الكلاب مما يؤدي إلى انتشار الإصابة بالأكياس العذارية .

نشكل هذة الدراسة أساسا لأي دراسات مستقبلية وخاصة الدراسات المورفولوجيه للدودة البالغة في العوائل الأساسية وأهميتها في أنها قادرة على تحديد أشكال استمرار دورة الحياة فصفات اليرقة والدودة البالغة متشابهة بالإضافة إلى الدراسة البيولوجيه الجزيئية وخاصة دراسة الحامض النوري DNA لميتوكوندريا الرويسات المعزولة من الأغنام والأبقار والإبل والإنسان

الهافلات تبد فعة الراء والهو للتامدية هي متن وتبليق فللودمو لاملد

G. S. P. L. A. J.

AL TAHDI UNIVERSITY الإقع الانشاري . لئع/26/1/2600 ف



وماهرية المربرة البيبة الشميرة الإشاراكية المختمت

الأعطرال يوموات

كبلية العلوم القارين

المواقلة الإلساسة 7007 ف

كليسسة العلسسوم فسسم الأحيسساء

منسبوان المحسبث

((مراسة انتشار و الشكل الخارجي للطور اليرقي للمشوكة المبيبية في الميوانات الهاشبة اللاليفية في منطقة سرت –ليبيا))

> مقدمسية مستنور الطبيسالي عبد القادر ممسد عبد القادر مفتسام

> > * * لجنــة المناقشــة :

الدكتور / حامست لحمرسدة قامسم (مشسرف الرسسالية)

الدكتور / فهرمسة حامسد همسام (معتصن داخلسس)

الدكتور / عبد الناصر على البوتي (ممتحین خارجیی)

حاسا الكين

06/12/4 - -

e frites



جامعة التحدي كليـــــة العلـــوم قســــم علـــم الحيـــــوان

دراسة انتشار والشكل الخارجي للطور اليرقى للمشوكة الحبيبية في الحيوانات العاشبه الأليفة في منطقة سرت ـ ليبيا

أطروحة مقدمة كجزء من متطلبات الأجازة العليا (الماجمىتير) في العلوم

مقدمة من الطالب عبد القادر مدمسد عبد القادر مدمست

أشراف الدكتور حامــــد احميـــدة قاســـم أستــــاذ مشــــارك _ علـــم الطفــيايات كلية العلوم _ جامعة قاريونــس بنغـــــازي _ لـــيـــا

ديممير (الكاتون) – 2006



Al-Tahaddi University

Faculty of Science
Department of Computer science
Sirite - G. S. P. L. A. J.

New Implementation of Unsupervised ID3 Algorithm (NIU-ID3)

Using Visual Basic.net

By:
Ahmed Ali Mohammed Alhouni

Supervisor: Dr. Faraj A. El-Mouadib

A thesis submitted to the department of computer science in partial fulfillment of the requirements for the degree of

Master of Science

Academic year 2008-2009